PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6 :
A01H 5/00, C12N 1/00, 1/21, 5/14, 15/29, 15/52, 15/70, 15/74, 15/82, C12Q 1/68

A1

(11) International Publication Number: WO 00/08920

(43) International Publication Date: 24 February 2000 (24.02.00)

(21) International Application Number: PCT/US99/18327

(22) International Filing Date: 12 August 1999 (12.08.99)

(30) Priority Data:
09/134,607 14 August 1998 (14.08.98) US

(71) Applicant *(for all designated States except US)*: YISSUM RESEARCH AND DEVELOPMENT COMPANY OF THE HEBREW UNIVERSITY OF JERUSALEM [IL/IL]; 46 Jabotinsky Street, 91042 Jerusalem (IL).

(71)(72) Applicant and Inventor *(for TJ only)*: FRIEDMAN, Mark, M. [US/IL]; Alharizi 1, 43406 Raanana (IL).

(72) Inventors; and
(75) Inventors/Applicants *(for US only)*: HIRSCHBERG, Joseph [IL/IL]; Burla 26, 93710 Jerusalem (IL). RONEN, Gil [IL/IL]; Mivtza Danny 39, 84456 Beer Sheva (IL). ZAMIR, Danny [IL/IL]; Derech Eretz 10, 70700 Gedera (IL).

(74) Common Representative: FRIEDMAN, Mark, M.; c/o Castorina, Anthony, Suite 207, 2001 Jefferson Davis Highway, Arlington, VA 22202 (IL).

(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published
With international search report.

(54) Title: POLYNUCLEOTIDES CONTROLLING THE EXPRESSION OF AND CODING FOR GENE B IN TOMATO

(57) Abstract

An isolated complementary or genomic DNA segment encoding lycopene cyclase of the B locus of tomato and its control elements which are responsible for its transient expression in chromogenic tissues such as fruit and flower are provided.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	**ES**	Spain	**LS**	Lesotho	**SI**	Slovenia
AM	Armenia	**FI**	Finland	**LT**	Lithuania	**SK**	Slovakia
AT	Austria	**FR**	France	**LU**	Luxembourg	**SN**	Senegal
AU	Australia	**GA**	Gabon	**LV**	Latvia	**SZ**	Swaziland
AZ	Azerbaijan	**GB**	United Kingdom	**MC**	Monaco	**TD**	Chad
BA	Bosnia and Herzegovina	**GE**	Georgia	**MD**	Republic of Moldova	**TG**	Togo
BB	Barbados	**GH**	Ghana	**MG**	Madagascar	**TJ**	Tajikistan
BE	Belgium	**GN**	Guinea	**MK**	The former Yugoslav Republic of Macedonia	**TM**	Turkmenistan
BF	Burkina Faso	**GR**	Greece			**TR**	Turkey
BG	Bulgaria	**HU**	Hungary	**ML**	Mali	**TT**	Trinidad and Tobago
BJ	Benin	**IE**	Ireland	**MN**	Mongolia	**UA**	Ukraine
BR	Brazil	**IL**	Israel	**MR**	Mauritania	**UG**	Uganda
BY	Belarus	**IS**	Iceland	**MW**	Malawi	**US**	United States of America
CA	Canada	**IT**	Italy	**MX**	Mexico	**UZ**	Uzbekistan
CF	Central African Republic	**JP**	Japan	**NE**	Niger	**VN**	Viet Nam
CG	Congo	**KE**	Kenya	**NL**	Netherlands	**YU**	Yugoslavia
CH	Switzerland	**KG**	Kyrgyzstan	**NO**	Norway	**ZW**	Zimbabwe
CI	Côte d'Ivoire	**KP**	Democratic People's Republic of Korea	**NZ**	New Zealand		
CM	Cameroon			**PL**	Poland		
CN	China	**KR**	Republic of Korea	**PT**	Portugal		
CU	Cuba	**KZ**	Kazakstan	**RO**	Romania		
CZ	Czech Republic	**LC**	Saint Lucia	**RU**	Russian Federation		
DE	Germany	**LI**	Liechtenstein	**SD**	Sudan		
DK	Denmark	**LK**	Sri Lanka	**SE**	Sweden		
EE	Estonia	**LR**	Liberia	**SG**	Singapore		

POLYNUCLEOTIDES CONTROLLING THE EXPRESSION OF AND CODING FOR GENE B IN TOMATO

FIELD AND BACKGROUND OF THE INVENTION

The present invention relates to a novel polynucleotide sequences isolated from tomato and, more particularly, to a novel lycopene cyclase gene and novel control elements controlling its specific expression in chromogenic tissues of plants, e.g., fruit and flower.

Carotenoids - functions and biosynthesis: Carotenoids comprise one of the largest classes of pigments in nature. In photosynthetic organisms carotenoids serve two major functions - as accessory pigments for light harvesting, and as protective agents against photooxidation processes in the photosynthetic apparatus. Another important role of carotenoids in plants, as well as in some animals, is that of providing distinctive pigmentation. Most of the orange, yellow, or red colors found in the flowers, fruits and other organs of many higher plant species are due to accumulation of carotenoids in the cells.

The biosynthesis of carotenoids has been reviewed extensively (Britton, 1988; Sandmann, 1994a). Carotenoids are produced from the general isoprenoid biosynthetic pathway, which in plants takes place in the chloroplasts of photosynthetic tissues and chromoplasts of fruits and flowers.

The first unique step in carotenoid biosynthesis is the head-to-head condensation of two molecules of geranylgeranyl pyrophosphate (GGPP) to produce phytoene (Figure 1). All the subsequent steps in the pathway occur in association with membranes. Four desaturation (dehydrogenation) reactions convert phytoene to lycopene via phytofluene, ζ-carotene, and neurosporene, as intermediates. Two cyclization reactions convert lycopene to β-carotene (Figure 1). Further reactions involve the addition of various oxygen-containing side groups which form the various xanthophyll species (not shown).

It has been established in recent years that four enzymes in plants catalyze the biosynthesis of β-carotene from GGPP: phytoene synthase, phytoene desaturase, ζ-carotene desaturase and lycopene cyclase (reviewed in Sandmann, 1994b). All enzymes in the pathway are nuclear encoded. Genes for phytoene synthase and phytoene desaturase have been previously cloned from tomato (Ray et al., 1992; Pecker et al., 1992).

The red color of ripe tomatoes is provided by lycopene, a linear carotene which accumulates during fruit ripening as membrane-bound crystals in chromoplasts (Laval-Martin et al., 1975). It is presumed to serve as an attractant of predators that eat the fruit and disperse the seeds. Accumulation of lycopene begins at the "breaker" stage of fruit ripening after the fruit has reached the "mature green" stage. In the "breaker" stage, which is indicated by the commencement of color change from green to orange, chlorophyll is degraded and chloroplasts turn into chromoplasts (Gillaspy et al., 1993; Grierson and Schuch, 1993). Total carotenoid concentration increases between 10 to 15-fold during the transition from "mature green" to "red". This change is due mainly to a 300-fold increase in lycopene (Fraser et al., 1994).

The cDNA which encodes lycopene β-cyclase, *CrtL-b*, was cloned from tomato (*Lycopersicon esculentum* cv. VF36) and tobacco (*Nicotiana tabacum* cv. Samsun NN, Pecker et al., 1996, U.S. Pat. application No. 08/399,561 and PCT/US96/03044 (WO 96/28014) both are incorporated by reference as if fully set forth herein) and was functionally expressed in *Escherichia coli*. This enzyme converts lycopene to β-carotene by catalyzing the formation of two β-rings, one at each end of the linear carotene. The enzyme interacts with half of the carotenoid molecule and requires a double bond at the C-7,8 (or C-7,8') position. Inhibition experiments in *E. coli* indicated that lycopene cyclase is the target site for the inhibitor 2-(4-methylphenoxy)tri-ethylamine hydrochloride (MPTA, Pecker et al., 1996). The primary structure of lycopene cyclase in higher plants is significantly conserved with the enzyme from cyanobacteria but differs from that of the non-photosynthetic bacteria *Erwinia* (Pecker et al., 1996). Levels of mRNAs of *CrtL-b* and *Pds*, which encodes phytoene desaturase, were measured in leaves, flowers and ripening fruits of tomato. In contrast to genes that encode enzymes of early steps in the carotenoid biosynthesis pathway, whose transcription increases during the "breaker" stage of fruit ripening, the level of *CrtL-b* mRNA decreases at this stage (Pecker et al., 1996). Hence, the accumulation of lycopene in tomato fruits is apparently due to a down-regulation of the lycopene cyclase gene that occurs at the breaker stage of fruit development. This conclusion supports the hypothesis that transcriptional regulation of gene expression is a predominant mechanism of regulating carotenogenesis.

The search for tissue specific control elements in plants is on going, however, only limited number of tissue specific control elements capable of

specifically directing gene expression in chromogenic tissues (fruit, flower) have so far been isolated. These include the promoters of the genes E4 and E8 (Montgomery et al., 1993), which are up-regulated by increase in ethylene concentration during tomato fruit ripening, the tomato gene 2A11 gene (Van Haaren and Houck, 1991) and the polygalacturonase (PG) gene (Nicholass et al., 1995; Montgomery et al., 1993), which are upregulated in tomato fruits during ripening.

There is thus a widely recognized need for, and it would be highly advantageous to have, a novel tissue specific control elements capable of specifically directing gene expression in chromogenic tissues.

The search for structural genes encoding enzymes associated with carotenogenesis is ongoing, and every new gene isolated not only provides insight into carotenogenesis, but also provides a tool to control and modify carotenogenesis for commercial purposes (Hirschberg et al. 1997, Cunningham FX Jr. and Gantt B, 1998).

There is thus a widely recognized need for, and it would be highly advantageous to have, a novel lycopene cyclase capable of altering the composition of carotenoids in carotenoids producing organisms.

SUMMARY OF THE INVENTION

According to one aspect of the present invention there is provided an isolated complementary or genomic DNA segment comprising a nucleotide sequence coding for a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, with the provision that the polypeptide has a major lycopene cyclase catalytic activity.

According to further features in preferred embodiments of the invention described below, the nucleotide sequence is selected from the group consisting of SEQ ID NOs: 8, 9, 10 and 11 and functional naturally occurring and man-induced variants thereof.

According to still further features in the described preferred embodiments the nucleotide sequence is a cDNA or a genomic DNA isolated form tomato.

According to another aspect of the present invention there is provided a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, the polypeptide having a major lycopene cyclase catalytic activity.

According to another aspect of the present invention there is provided a transduced cell overexpressing a polypeptide including an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, the polypeptide having a major lycopene cyclase catalytic activity, the cell therefore over producing β-carotene on an expense of lycopene.

According to still further features in the described preferred embodiments the transduced cell is selected from the group consisting of a prokaryotic cell and a eukaryotic cell.

According to still further features in the described preferred embodiments the eukaryotic cell is of a higher plant.

According to still further features in the described preferred embodiments the cell forms a part of a transgenic plant.

According to yet another aspect of the present invention there is provided a method of down-regulating production of β-carotene in a cell comprising the step of introducing into the cell at least one anti-sense polynucleotide sequence capable of base pairing with messenger RNA coding for a polypeptide including an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, the polypeptide having a major lycopene cyclase catalytic activity, the cell therefore under producing β-carotene from lycopene.

According to still further features in the described preferred embodiments the at least one anti-sense polynucleotide sequence includes a synthetic oligonucleotide.

According to still further features in the described preferred embodiments the synthetic oligonucleotide includes a man-made modification rendering the synthetic oligonucleotide more stable in cell environment.

According to still further features in the described preferred embodiments the synthetic oligonucleotide is selected from the group consisting of methylphosphonate oligonucleotide, monothiophosphate oligonucleotide, dithiophosphate oligonucleotide, phosphoramidate oligonucleotide, phosphate ester oligonucleotide, bridged phosphorothioate oligonucleotide, bridged phosphoramidate oligonucleotide, bridged methylenephosphonate oligonucleotide, dephospho internucleotide analogs with siloxane bridges, carbonate bridge oligonucleotide, carboxymethyl ester bridge oligonucleotide, carbonate bridge oligonucleotide,

carboxymethyl ester bridge oligonucleotide, acetamide bridge oligonucleotide, carbamate bridge oligonucleotide, thioether bridge oligonucleotide, sulfoxy bridge oligonucleotide, sulfono bridge oligonucleotide and α-anomeric bridge oligonucleotide.

According to still further features in the described preferred embodiments the at least one anti-sense polynucleotide sequence is encoded by an expression vector.

According to still further features in the described preferred embodiments the cell is selected from the group consisting of a prokaryotic cell and a eukaryotic cell.

According to still further features in the described preferred embodiments the eukaryotic cell is of a higher plant.

According to still further features in the described preferred embodiments the cell forms a part of a transgenic plant.

According to still another aspect of the present invention there is provided an expression construct for directing an expression of a gene in fruit or flower comprising a regulatory sequence selected from the group consisting of an upstream region of a B allele of tomato and an upstream region of a b allele of tomato.

According to still further features in the described preferred embodiments the expression construct comprising a functional part of nucleotides 1-1210 of SEQ ID NO: 14 or nucleotides 1-1600 of SEQ ID NO: 15, or functional naturally occurring and man-induced variants thereof.

According to still further features in the described preferred embodiments the expression construct comprising at least one control element having a sequence selected from the group consisting of SEQ ID NOs:21-24, all derived from SEQ ID NO:11, and functional naturally occurring and man-induced variants thereof.

According to still further features in the described preferred embodiments the expression construct is selected from the group consisting of plasmid, cosmid, phage, virus, bacmid and artificial chromosome.

According to still further features in the described preferred embodiments the expression construct is designed to integrate into a genome of a host.

According to yet another aspect of the present invention there is provided a transduced cell or transgenic plant transduced with the above described expression construct.

According to still another aspect of the present invention there is provided a method of isolating a gene encoding a polypeptide having an amino acid sequence homologous to SEQ ID NOs: 17, 18 and 19 and having a major lycopene cyclase catalytic activity from a species, the method comprising the step of screening a complementary or genomic DNA library prepared from isolated RNA or genomic DNA extracted from the species with a probe having a sequence derived from SEQ ID NOs: 8, 9, 10 or 11 and isolating clones reacting with the probe.

The present invention successfully addresses the shortcomings of the presently known configurations by providing novel polynucleotides controlling the expression of genes in fruit and flower in plant and a novel polynucleotide encoding lycopene cyclase.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention herein described, by way of example only, with reference to the accompanying drawings, wherein:

FIG. 1 presents the pathway of carotenoid biosynthesis in plants and algae. Enzymes are indicated by the their gene assignment symbols: *aba2*, zeaxanthin epoxidase; *CrtL-b*, Lycopene β-cyclase; *CrtL-e*, lycopene ε-cyclase; *CrtR-b*, β-ring hydroxylase; *CrtR-e*, ε-ring hydroxylase; *Pds*, phytoene desaturase (*crtP* in cyanobacteria); *Psy*, phytoene synthase (*crtB* in cyanobacteria); *Zds*, ζ-carotene desaturase (*crtQ*) in cyanobacteria. GGDP, geranylgeranyl diphosphate.

FIG. 2 shows fine genetic mapping and molecular organization of *B* on chromosome 6 of the tomato linkage map. The linkage map was adopted from Eshed and Zamir (1995). The relevant chromosomal segments from *L. pennellii* that were introgressed to *L. esculentum* lines IL 6-2 and IL 6-3 are represented by black bars. High-resolution genetic map around *B* is displayed with genetic distances in map units (cM). Positions of the YAC inserts are designated under the map.

FIG. 3 demonstrates levels of mRNA (relative units) during fruit ripening of wild-type tomato *L. esculentum* . Data are derived from quantifying the DNA products in the RT-PCR analysis of total RNA extracted at different stages of fruit development. Ripening stages: IG, immature green; MG, mature green, B, breaker, O, Orange; P, pink; R, red.

FIG. 4 demonstrates levels of mRNA (relative units) during fruit ripening of the tomato mutant *High-beta*. Data are derived from quantifying the DNA products in the RT-PCR analysis of total RNA

extracted at different stages of fruit development. Ripening stages: G, green; MG, mature green, B, breaker, O, Orange; P, pink; R, red.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is of novel polynucleotide sequences isolated from tomato which can be used to control gene expression in plant chromogenic tissues, especially fruit and flower. The present invention is further of polynucleotide sequences isolated from tomato which encode a lycopene cyclase which can be used to alter carotenogenesis is carotenoids producing organisms.

The principles and operation of the present invention may be better understood with reference to the drawings and accompanying descriptions.

Before explaining at least one embodiment of the invention in detail, it is to be understood that the invention is not limited in its application to the details of construction and the arrangement of the components set forth in the following description or illustrated in the drawings. The invention is capable of other embodiments or of being practiced or carried out in various ways. Also, it is to be understood that the phraseology and terminology employed herein is for the purpose of description and should not be regarded as limiting.

Fruit of the cultivated tomato (*Lycopersicon esculentum*) accumulate lycopene, a red carotenoid pigment. A dominant allele of gene *B* determines accumulation of β-carotene in the fruits of the tomato mutant '*high-beta*', at the expense of lycopene, resulting in a unique orange color. Conversion of lycopene to β-carotene in the biosynthesis pathway of carotenoids is catalyzed by the enzyme lycopene β-cyclase. Previously it was shown that *CrtL-b*, the gene for lycopene β-cyclase, does not map to the locus *B* in the tomato genetic map. This ruled out the possibility that a mutation in lycopene β-cyclase encoded by *CrtL-b* causes the phenotype in *high-beta*.

The locus *B* was mapped to chromosome No. 6. The dominant allele *B* was found in the tomato introgression line IL 6-2. The DNA of *B* was identified and cloned by a map-based (positional) cloning method. The nucleotide sequence of this gene was determined and demonstrated a novel type of a lycopene cyclase enzyme. Its primary structure has some similarity to other lycopene cyclases and to the enzyme capsanthin-capsorubin synthase from pepper. In addition, nucleotide sequence was

identified, which functions as a strong promoter during fruit development in the *B* allele of the mutant *High-beta*.

Thus, according to one aspect of the present invention there is provided an isolated complementary or genomic DNA segment comprising a nucleotide sequence coding for a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof. The polypeptide has a major lycopene cyclase catalytic activity. Polypeptides which share at least 70, 75, 80, 85, 90, 95 or more identical amino acid residues with SEQ ID NOs: 17, 18 or 19 are also within the scope of the present invention.

As used herein in the specification and in the claims section below, the phrase "major lycopene cyclase catalytic activity" refers to catalytic activity mainly directed at the conversion of lycopene to β-carotene by catalyzing the formation of two β-rings, one at each end of the linear carotene, such that if introduced into lycopene-accumulating *E. coli* cells, such cells accumulate also β-carotene up to a range of at least few percent e.g., 5 %, to preferably about 15 %, or more, of total carotenoids therein by symmetric formation of two β-ionone rings on the linear lycopene molecules therein.

According to a preferred embodiment of the invention the nucleotide sequence is as set forth in SEQ ID NOs: 8, 9, 10 or 11, or functional naturally occurring or man-induced variants thereof. As further shown below these sequences are genomic and complementary DNA sequences which were derived while reducing the present invention to practice from certain tomato cultivars or lines. However, nucleotide sequences which share 70, 75, 80, 85, 90, 95 or more identical nucleotides with SEQ ID NOs: 8, 9, 10 or 11 are also within the scope of the present invention.

According to another aspect of the present invention there is provided a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, the polypeptide having a major lycopene cyclase catalytic activity. Homologous polypeptides as describe above and further detailed hereinunder are also envisaged.

According to another aspect of the present invention there is provided a transduced cell overexpressing a polypeptide including an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19, and functional naturally occurring and man-induced variants

thereof, the polypeptide having a major lycopene cyclase catalytic activity, the cell therefore over producing β-carotene on an expense of lycopene.

The cell according to the present invention can be of any type. For example, the cell can be a prokaryotic cell or a eukaryotic cell. Preferably the cell is of a higher plant. The cell preferably forms a part of a transgenic plant. Methods of transducing cells (and cells in organisms to form transgenic organisms) are well known in the art and do not require further description herein. Protocols are available, for example, in (Sambrook et al., 1989).

As used herein in the specification and in the claims section below, the term "transduced" refers to the result of a process of inserting nucleic acids into cells. The insertion may, for example, be effected by transformation, viral infection, injection, transfection, gene bombardment, electroporation or any other means effective in introducing nucleic acids into cells. Following transduction the nucleic acid is either integrated in all or part, to the cell's genome (DNA), or remains external to the cell's genome, thereby providing stably transduced or transiently transduced cells.

According to yet another aspect of the present invention there is provided a method of down-regulating production of β-carotene in a cell comprising the step of introducing into the cell at least one anti-sense polynucleotide sequence capable of base pairing with messenger RNA coding for a polypeptide including an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, the polypeptide having a major lycopene cyclase catalytic activity, the cell therefore under producing β-carotene from lycopene. Again, the cell can be of any type. For example, the cell can be a prokaryotic cell or a eukaryotic cell. Preferably the cell is of a higher plant. The cell preferably forms a part of a transgenic plant.

As used herein in the specification and in the claims section below, the term "down regulating" means also reducing, lowering, inhibiting, etc., e.g., permanently or transiently reducing.

As used herein in the specification and in the claims section below, the term "production" means also formation or generation.

As used herein in the specification and in the claims section below, the term "introducing" means also providing with or inserting.

The at least one anti-sense polynucleotide sequence according to the present invention can includes one or several synthetic oligonucleotides capable of base pairing with messenger RNA derived from the above-

identified nucleotide sequences. The synthetic oligonucleotide preferably includes a man-made modification rendering the synthetic oligonucleotide more stable in cell environment. The modified oligonucleotide can be, for example, a methylphosphonate oligonucleotide, monothiophosphate oligonucleotide, dithiophosphate oligonucleotide, phosphoramidate oligonucleotide, phosphate ester oligonucleotide, bridged phosphorothioate oligonucleotide, bridged phosphoramidate oligonucleotide, bridged methylenephosphonate oligonucleotide, dephospho internucleotide analogs with siloxane bridges, carbonate bridge oligonucleotide, carboxymethyl ester bridge oligonucleotide, carbonate bridge oligonucleotide, carboxymethyl ester bridge oligonucleotide, acetamide bridge oligonucleotide, carbamate bridge oligonucleotide, thioether bridge oligonucleotide, sulfoxy bridge oligonucleotide, sulfono bridge oligonucleotide or an α-anomeric bridge oligonucleotide. For further details the reader is referred to Cook (1991).

Alternatively, the anti-sense polynucleotide sequence is encoded by an anti-sense expression vector. Such vectors are well known in the art and are commercially available from, for example, pBI101, pBI121, pBI221 (commercially available from Colntech.)

Further according to the present invention, there is provided an expression construct for directing an expression of a gene in fruit or flower of a plant. The expression vector according to the present invention includes a regulatory sequence selected from the group consisting of an upstream region of a B allele of tomato and an upstream region of a b allele of tomato. Thus, according to a preferred embodiment of the invention, the expression construct includes a functional part of nucleotides 1-1210 of SEQ ID NO: 14 or nucleotides 1-1600 of SEQ ID NO: 15, or functional naturally occurring and man-induced variants thereof.

According to a preferred embodiment, the expression construct includes at least one control element having a sequence selected from the group consisting of SEQ ID NOs: 21-24, all derived from SEQ ID NO:11, and functional naturally occurring and man-induced variants thereof.

As further detailed in the Examples section hereinbelow, these sequence elements, which are 26, 13, 9, and 8 bp long and start at (5' end) nucleotides 859, 753, 479 and 306, respectively, of SEQ ID NOs: 11, 15, are located upstream to the initiator methionine codon in the B allele are the main difference between the B and b allele, and are therefore responsible for the differential expression of the B locus in tomato.

The expression construct according to the present invention can be a plasmid, cosmid, phage, virus, bacmid or an artificial chromosome. Each of these constructs has unique sequences rendering the construct most applicable for some as opposed to other applications, as well known in the art. Regardless of its type, according to a preferred embodiment of the present invention the expression construct is designed to integrate into a genome of a host, such that stable transfectants are obtainable. However, the scope of the present invention is not limited to such constructs. In other words, constructs designed for transient transfection are also within the scope of the present invention. In any case, the construct preferably includes at least one positive and/or negative selection gene, and is suitable for transformation, transfection, transgenization and gene knock-in procedures.

According to yet another aspect of the present invention there is provided a transduced cell or a transgenic plant transduced with the above described expression construct. Such a cell or plant is expressing the gene located downstream to the regulatory sequence in a controlled developmental manner, mimicking the expression of the lycopene cyclase gene of the B locus in b or B tomato plants.

According to still another aspect of the present invention there is provided a method of isolating a gene encoding a polypeptide having an amino acid sequence homologous to SEQ ID NOs: 17, 18 and 19 and having a major lycopene cyclase catalytic activity from a species. The method is effected by executing the following method steps, in which a complementary or genomic DNA library prepared from isolated RNA or genomic DNA extracted from the species is screened with a probe having a sequence derived from SEQ ID NOs: 8, 9, 10 or 11 and clones reacting with the probe are isolated. Such clones are good candidates to include segments of genes homologous to SEQ ID NOs: 8, 9, 10 or 11, which genes are good candidates to encode a polypeptide having an amino acid sequence homologous to SEQ ID NOs: 17, 18 and 19. 5' cloning strategies, such as, but not limited to RACE protocols can be employed to isolate full length clones, as well known in the art.

Thus, according to the present invention, the following uses of gene B of tomato are anticipated:

(i) Increasing the content of β-carotene in tissues of transgenic plants over-expressing it. This is an advantageous attribute in fruits and

vegetables because it will provide better nutritional value and enhanced color.

(ii) Increasing the accumulation of lycopene in fruits and flowers of transgenic plants by reducing the activity of B using anti-sense inhibition, preferably via anti-sense expression.

(iii) Achieving strong expression of transgenes specifically in fruits and flowers using the promoter sequence of the gene B from *High-beta* tomato cultivars.

Each of the various and aspects of the present invention as delineated hereinabove and as claimed in the claims section below finds experimental support in the Examples section that follows.

EXAMPLES

Bacteria and plants: *E. coli* strain XL1-Blue was used in all experiments described herein. Tomato (*Lycopersicon esculentum*) CV M82 served as the 'wild-type' strain in the fruit ripening measurements. The introgression lines IL 6-2 and IL 6-3 (Eshed and Zamir, 1994) were used as a source for the *B* mutation and employed for fine mapping of the *B* locus.

Fine mapping and cloning of the B locus: As a source to B mutation, the lines IL-6-2 or IL-6-3 (*BB*) were used (Eshed and Zamir, 1995). Each line was crossed with the cultivated tomato cv M-82 (*bb*), and the hybrids were selfed to create an F-2 population that segregated for both the *B* phenotype and the introgressed DNA segment. 1335 F-2 plants were scored for the RFLP using markers CT193 and TG578 (Pnueli et al., 1998; Tanksley et al., 1992) and for the *B* phenotype, and recombinant plants were collected. The 32 resulting recombinants were further screened with all the available RFLP probes surrounding *B* to accurately map the mutated locus (Figure 2). One RFLP marker, TM16 (Pnueli et al., 1998), was co-segregated with *B* in less than 0.0375 cM resolution.

The tomato genomic library in YACs was screened with DNA of markers TM16 and TG275. Two overlapping YAC clones, designated 271 and 310, were identified by hybridization. DNA sequences from the ends of the inserts in these YACs were amplified by PCR as previously described (Pnueli et al., 1998) and were used as molecular probes to screen the 32 recombinant plants for Restriction Fragment Length Polymorphism (RFLP). The YAC ends were mapped as shown in Figure 2. It was established that YAC 310 overlaps the B locus, thus ensured that the 200 kb insert of YAC

310 contains the B gene. In contrast, recombination between the left end of YAC 271 (271le) and the B phenotype indicated that this YAC clone did not carry the B locus and defined its location in a relatively small region of YAC 310 that did not overlap with YAC 271 (Figure 2).

The DNA insert of YAC 310 was cut with EcoRI and the resulting fragments were subcloned in the vector λ-gt11. Two phage clones designated B1 and B3, co-segregated with the B locus and mapped to the end of YAC 310. The nucleotide sequence of the insert of B1 was determined. The B1 fragment was further used to screen a genomic library of wild-type tomato (cv VF36) in the lambda vector EMBL3, and a cosmid library of *L. pennellii*. A single positive phage clone and a single positive cosmid clone were isolated, respectively.

The B1 fragment was also used to screen 1.5 million plaques of a cDNA library from a tomato fruit and 3 identical clones were isolated. The ca. 1300 bp inserts in these clones contained an open reading frame that was lacking the 5' end, as determined by nucleotide sequence analysis. The full-length cDNAs were then obtained using reverse-transcription polymerase chain reaction (RT-PCR) method with RNA isolated from wild-type (VF-36) and *high-beta* (IL 6-3)flowers and fruits. For the PCR reaction we used 5' primers based on the genomic sequence taken from the sequence of B1 insert and the 3' primers based on the cloned cDNA. The full coding region of the cDNA of the allele b of wild type tomato (cv. VF-36) and the allele B from *L. pennellii* were excised in pBluescript KS- vector which were designated pBESC and pBPENN, respectively. DNA sequence comparison between cDNA and genomic sequences revealed no introns interference in the genomic sequence of the b (and B).

DNA blot hybridization was done according to conventional techniques (Sambrook et al., 1989, Eshed and Zamir, 1994) at low stringency in a buffer containing 10 x Denharts, 5 x SSC, 50 mM phosphate buffer (pH-7), 1 % SDS, 50 mg salmon sperm (sheared, autoclaved and boiled before adding to the mixture). Filters were washed with 5 x SSC at 65 °C.

Genomic DNA of tomato was prepared from 5 grams of leaf as previously described (Eshed and Zamir, 1995).

Amplification by the polymerase chain reaction (PCR) method of the full length cDNA of the b allele was carried out with the following oligonucleotide primers, whose sequence was derived from the genomic sequence of the B1 clone (see below): Forward: 5'-

AATGGAAGCTCTTCTCAAGCCT-3' (SEQ ID NO:1), Reverse: 5'-CACATTCAAAGGCTCTCTATCGC-3' (SEQ ID NO:2).

Total RNA was extracted from 1.5 grams of fruit or 0.1 gram of flower or leaf tissues as previously described (Pecker et al., 1996).

Measurement of mRNA levels by the reverse transcription followed by polymerase chain reaction (RT-PCR) technique was carried out as previously described (Pecker et al., 1996) using the following oligonucleotides as primers for the PCR reaction. For amplification of the gene Psy the following primer were employed: Forward1: 5'-TCGAGAACGGACGATG-3' (SEQ ID NO:3), Forward2 (internal): 5'-TGCAGAGAGACAGATG-3' (SEQ ID NO:4) and Reverse: 5'-ATTTCATGCTTTATCTTTGAAG-3' (SEQ ID NO:5).

For amplification of allele B: Forward 5'-GCTGAAGTTGAAATTGTTGA-3' (SEQ ID NO:6) and Reverse 5'-TCTCTTCCTCAATAACACTT-3' (SEQ ID NO:7).

Sequence analysis: DNA sequence analysis was performed by the ABI Prism 377 DNA sequencer (Perkin Elmer) and processed with the ABI sequence analysis software. Nucleotide and amino acid sequence analysis and comparisons were done using the UWGCG software package.

Plasmids: Plasmid pACCRT-EIB for expressing bacterial carotenoid biosynthesis genes in *E. coli*, was previously described (Cunningham et al., 1993). Plasmid pBESC and pBPENN were constructed by inserting an 1666 bp of cDNA of the tomato *B* allele (from *L. pennellii*) or *b* allele (from *L. esculentum*), respectively, in the EcoRV site of the plasmid vector pBluescript KS$^-$ (Stratagene®).

Pigment extraction and analysis: For extraction of pigments from *E. coli*, aliquots of 2 ml were taken from bacterial suspension cultures. The cells were harvested by centrifugation, washed once with water, resuspended in 2 ml of acetone and incubated at 65 °C for 10 minutes in the dark. The samples were centrifuged again at 13,000 *g* for 5 minutes and the acetone supernatant containing the pigments was placed in a clean tube. More than 99 % of the carotenoids were extracted by this procedure as determined by re-extraction after breaking and grinding the samples. The pigment extract was blown to dryness under a stream of N_2 and stored at -20 °C until required for analysis.

Fruit pigments were extracted from 1.0 gram of fresh tissue. The tissue was ground in 2 ml of acetone and incubated at room temperature in the dark for 10 minutes. Then, 2 ml of dichloro-methane were added and

the samples were agitated until all pigments were transferred to the supernatant, which was then filtered. To each sample, 4 ml of ether and 0.4 ml of 12 % w/v $NaCl/H_2O$ were added and the mixture was shaken gently until all pigment was transferred to the upper (ether) phase. The ether was collected, and the pigment extract was blown to dryness under a stream of N_2 and stored at -20 °C until required for analysis.

Carotenoids were separated by reverse phase HPLC using a Spherisorb ODS-2 column (silica 5 mm 3.2 mm x 250 mm, Phenomenex®). Samples of 50 µl of acetone-dissolved pigments were injected to a Waters 600 pump. The mobile phase consisted of acetonitrile:H_2O (9:1) - solvent A, and 100 % ethyl acetate - solvent B, which were used in a linear gradient between A and B for 30 minutes, at flow of 1 ml per minute. Light absorption peaks were detected in the range of 200-600 nm using a Waters 996 photo diode-array detector. All spectra were recorded in the eluting HPLC solvent, as was the fine absorbance spectral structure. Carotenoids were identified by their characteristic absorption spectra and their typical retention time, which corresponded to standard compounds of lycopene and β-carotene. Peak areas were integrated by the Millennium chromatography software (Waters).

EXPERIMENTAL RESULTS

The only difference between the *high-beta* mutant and the wild-type tomato is in the fruit color due to accumulation of β-carotene at the expense of lycopene. Thus, it was logical to assume that this mutation occurred in the gene that encodes lycopene-β-cyclase (*CrtL-b*). However, the *CrtL-b* cDNA that was previously cloned from tomato (Pecker et al., 1996) was mapped to 2 loci on chromosomes Nos. 4 and 10, but not on chromosome 6, where the B locus was mapped. Even at very low stringency of hybridization conditions we were unable to detect any hybridization of the tomato *CrtL-b* like sequences on chromosome 6.

Therefore, the only way to clone the gene B, which is responsible for the high-beta phenotype, was to use map-based ("positional") cloning techniques.

Fine mapping of the B locus: As a source to the B mutation, the IL-6-2 or IL-6-3 (*BB*) (Eshed and Zamir, 1995) tomato lines were employed. Each line was crossed with the cultivated tomato cv. M-82 (*bb*), and the hybrids were selfed to create an F-2 population that segregated for both the

B phenotype and the introgressed DNA segment. 1335 F-2 plants were scored for the RFLP using markers CT-193 and TG-578, (Pnueli et al., 1998; Tanksley et al., 1992) and for the *B* phenotype, and recombinant plants were collected. The 32 recombinants collected were further screened with all the available RFLP probes surrounding *B* to accurately map the mutated locus (Figure 2). One RFLP marker, TM-16 (Pnueli et al., 1998), co-segregated with *B* in less than 0.0375 cM resolution.

The tomato genomic library in YACs was screened with the DNA marker TM-16 as a molecular probe. Two YAC clones, designated 271 and 310, were identified by hybridization. DNA sequences from the ends of the inserts in these YACs were amplified by PCR as previously described (Pnueli et al., 1998) and were used as molecular probes to screen the 32 recombinant plants for Restriction Fragment Length Polymorphism (RFLP). The YAC ends were mapped as shown in Figure 2. It was established that YAC 310 overlaps the B locus, thus ensured that the 200 kb insert of YAC 310 contains the B gene. In contrast, recombination between YAC 271 and the B phenotype indicated that this clone did not carry the B locus. Moreover, it established that B was residing in a confined small region of YAC 310 that did not overlap with YAC 271 (Figure 2).

The DNA insert of YAC 310 was cut with EcoRI and the resulting fragments were subcloned in the vector λ-gt11. Two phage clones designated B1 and B3, co-segregated with the B locus and mapped to the end of YAC 310. The nucleotide sequence of the insert of B1 was determined. The B1 fragment was further used to screen a genomic library of wild-type tomato (cv VF36) in the lambda vector EMBL3, and a cosmid library of *L. pennellii*. A single positive phage clone and a single positive cosmid clone were isolated, respectively.

The B1 fragment was also used to screen 1.5 million plaques of cDNA library from a tomato fruit and 3 identical clones were isolated. The ca. 1300 bp inserts in these clones contained an open reading frame that was lacking the 5' end, as determined by nucleotide sequence analysis. The full-length cDNAs were then obtained using reverse-transcription polymerase chain reaction (RT-PCR) method with RNA isolated from wild-type (VF-36) and *high-beta* (IL 6-3) flowers and fruits. For the PCR reaction we used 5' primers based on the genomic sequence taken from the sequence of B1 insert and the 3' primers based on the cloned cDNA. The full coding region of the cDNA of the allele b of wild type tomato (cv. VF-36) and the allele B from *L. pennellii* were excised in pBluescript KS$^-$ vector which were

designated pBESC and pBPENN, respectively. DNA sequence comparison between cDNA and genomic sequences revealed no introns interference in the cDNA sequence.

Table 1 below summarizes the sequence data with reference to the sequence listing:

TABLE 1

Type	allele	Species	SEQ ID NO:
cDNA	b	*L. esculentum*	8
gDNA	b	*L. esculentum*	9
cDNA	B	*L. pennellii*	10
gDNA	B	*L. pennellii*	11
cDNA	ogC	*L. esculentum*	12
translated cDNA	b/B	*L. esculentum* / *L. pennellii*	13
translated gDNA	b	*L. esculentum*	14
translated gDNA	B	*L. pennellii*	15
translated cDNA	ogC	*L. pennellii*	16
peptide (translated from cDNA)	b	*L. esculentum*	17
peptide (translated from gDNA)	b	*L. esculentum*	18
peptide (translated from cDNA)	B	*L. pennellii*	19
peptide (translated from cDNA)	ogC	*L. esculentum*	20

cDNA = complementary DNA; gDNA = genomic DNA; bp = base pairs; aa = amino acid.

Cloning and sequence analysis of old-gold-crimson (ogC) mutation: The old-gold and crimson are two names given to a well-known recessive mutation that was found in the Philippines in 1951 (Butler, 1962 and the SolGenes databases: http:// probe.nal.usda.gov:8300/ cgi-in/webace?db = solgenes & class = Locus & object = og; and: http:// probe.nal.usda.gov:8300/ cgi-bin/webace?db = solgenes & class = Image & object = og%2c + old + gold). The ogC locus was mapped to chromosome 6. At least 2000 F-2 progenies of a cross between *High-beta* (BB) and ogC were screened for B-ogC double mutants and not a single recombinant plant was found. That locates B and ogC less than 0.025 cM apart. The ogC phenotype is characterized by over accumulation of lycopene, both in fruits

and flowers, compare to wild type tomatoes and lack of β-carotene in the fruits.

Cloning the B locus from ogC mutant plants was done by PCR method on total genomic DNA extracted from ogC plants using primers that were based on the sequence of the b allele described herein. Sequence analysis of the b-homolog revealed a single base deletion, in the coding sequence of b at position 104 from the initiation codon (compare SEQ ID NOs: 13 and 16). This deletion created a frame-shift mutation that shortened the translatable polypeptide to 56 amino acids. This finding indicates that the ogC is a null mutation of the normal function of the b gene.

Sequences comparison of alleles in the B locus: Nucleotide sequence analysis of the 1666 bp cDNA revealed an open reading frame of 498 codons, potentially coding for a polypeptide of 498 amino acids with a calculated molecular mass of 56.4 kDa. Nucleotide sequence analysis showed 98% identity between b (from VF-36, SEQ ID NO: 8) and B (from *L. pennellii*, SEQ ID NO: 10). The amino acid sequences of B and b are 97.4% identical (SEQ ID NOs: 17 and 19).

In the 1200 bp sequences upstream to the translated region of B from *L. pennellii* there are four sequence insertions as compared with the equivalent region in b from VF-36. The inserts are 26, 13, 9, and 8 bp long and start at (5' end) nucleotides 859, 753, 479 and 306, respectively, of SEQ ID NOs: 11, 15. They are located upstream to the initiator methionine codon in the B allele are the main difference between the B and b alleles, and are therefore responsible for the differential expression of the B locus in tomato. Their sequences are TGACTTCACCCTTCTTTCTTGTCTTC (SEQ ID NO:21), AGAGTCTGGGTTC (SEQ ID NO:22), CTAGTATCG (SEQ ID NO:23) and CTAAATAT (SEQ ID NO:24). An additional AATTTTCAAA (SEQ ID NO:25) sequence, which is found in upstream regions of ethylene-activated genes such as E4 and E8 (Montgomery et al., 1993), is shared by the upstream regions of the B and b alleles. All other sequences in the promoter and region are 90-94% conserved in the two allele (compare SEQ ID NOs: 9 and 11).

The polypeptide products of B and b are β-carotene synthases: The use of *E. coli* heterologous system for carotenoid biosynthesis has been proven to be a powerful tool for identifying genes associated with carotenoid biosynthesis. *E. coli* cells of the strain XLI- Blue, carrying the plasmid pACCRT-EIB accumulate lycopene (Cunnungham et al. 1993).

Lycopene-accumulating *E. coli* cells were co-transformed with the plasmid pBESC or pBPENN and selected on LB medium containing both ampicillin and chloramphenicol. Carotenoids from cells carrying pACCRT-EIB alone, or pACCRT-EIB and either pBESC or pBPENN were extracted and analyzed by HPLC.

Cells carrying only the pACCRT-EIB plasmid produced lycopene, while cells carrying both pACCRT-EIB and pBPENN accumulate also β-carotene up to 13 % of total carotenoids. Similarly, cells carrying both pACCRT-EIB and pBESC produced β-carotene up to 5 % of total carotenoids (see Table 2 below). These results indicated that the cDNA-products of both the B and b alleles are capable of converting lycopene to β-carotene by the symmetric formation of two β-ionone rings on the linear lycopene molecule.

TABLE 2

The B gene product converts lycopene to β-carotene. **Accumulation of carotenoids in *E. coli* cells expressing alleles *B* or *b* from tomato (percent of total carotenoids)**

plasmid	lycopene	β-carotene
pACCRT-EIB	100	
pACCRT-EIB + pBESC (b)	87	13
pACCRT-EIB + pBPENN (B)	95	5

Sequence comparison between B and other carotene cyclases: The nucleotide sequences of the coding region of b and the coding region of the cDNA of the previously published lycopene β-cyclase from tomato, *CrtL-b* (Pecker et al, 1996) , are 59 % identical. The polypeptide products of these genes are only 52 % identical. These data explain why *CrtL-b* could not hybridize with the sequence of B. Moreover, while the similarity in amino acid sequence between B and CRTLB suggests a common mechanism of lycopene cyclization, it clearly demonstrates that B is a novel lycopene β-

cyclase enzyme. There is no similarity (less than 45 % identities) in the non-translated regions of these two genes.

Surprisingly, the nucleotide sequence of the cDNA of b is 83% identical with the cDNA of a gene from bell pepper (*Capsicum annuum*), which catalyzes the conversion of the ubiquitous 5,6-epoxycarotenoids, antheraxanthin and violaxanthin, into the ketocarotenoids capsanthin and capsorubin, respectively (Bouvier et al., 1994). This enzyme, called also capsanthin-capsorubin synthase (CCS), is synthesized specifically in pepper fruits. There is 85 % identity in the deduced amino acid sequences of B and CCS.

Expression of B gene during fruit ripening in wild-type and High-beta: Previously, it has been shown that the steady-state levels of mRNA of the genes for early enzymes in the carotenoid biosynthesis pathway, phytoene synthase and phytoene desaturase, increase during fruit ripening in tomato (Hirschberg et al., 1997). In the case of Pds it was demonstrated that transcriptional up-regulation is responsible for this increase (reviewed in Hirschberg et al., 1997). Recently, we have determined that the mRNA level of *CrtL-b*, which encodes lycopene β-cyclase, decreases during tomato fruit ripening (Pecker et al. 1996).

To determine the regulation of expression of B gene during fruit development in tomato, we have measured by RT-PCR its mRNA level at different stages of fruit development. As can be seen in Figure 3, mRNA of the b gene is undetected in leaves and during the green stages of fruit ripening of wild-type tomato. However, it is increased at the 'breaker' stage of ripening but then disappears at later stages of ripening. This marked drop of mRNA of B is contrasted by the dramatic increase in mRNA level of Psy at the same stages of fruit ripening.

In contrast to the wild-type tomato, the mRNA level of B in the fruit of the *High-beta* mutant (containing the B allele) increases dramatically at the 'breaker' stage and remains high during all the subsequent ripening stages (Figure 4). These results indicate that the major difference between alleles b and B is in the level of expression at different ripening stages. The results further explain the phenotype of mutant *High-beta*, carrying the B allele, where a novel type of lycopene cyclase, which is capable of converting lycopene to β-carotene, is highly expressed during fruit ripening.

Although the invention has been described in conjunction with specific embodiments thereof, it is evident that many alternatives,

modifications and variations will be apparent to those skilled in the art. Accordingly, it is intended to embrace all such alternatives, modifications and variations that fall within the spirit and broad scope of the appended claims.

REFERENCES CITED IN ALPHABETIC ORDER

1. Bouvier, F., Hugueney, P., d'Harlingue, A., Kuntz, M., and Camara, B. (1994). Xanthophyll biosynthesis in chromoplasts: Isolation and molecular cloning of an enzyme catalyzing the conversion of 5,6-epoxycarotenoid into ketocarotenoid. Plant J. 6:45-54.
2. Britton G (1988) Biosynthesis of carotenoids, Plant Pigments. TW Goodwin, ed, Academic Press), London and New York: pp. 133-180.
3. Butler L, (1962) A new fruit color, Rep. Tomato Genetic Cooperation 12:17-18.
4. Cunningham, F.X.,Jr., Chamovitz, D., Misawa, N., Gantt, E., and Hirschberg, J. (1993). Cloning and functional expression in Escherichia coli of a cyanobacterial gene for lycopene cyclase, the enzyme that catalyzes the biosynthesis of β-carotene. FEBS Lett. 328:130-138.
5. Cunningham F.X.,Jr. and Gantt, E. (1998) Genes and enzymes of carotenoid biosynthesis in plants. Annu. Rev. Plant Physiol. Plant Mol. Biol. 49:557-583.
6. Cook (1991) Medicinal chemistry of antisense oligonucleotides - future opportunities. Anti-Cancer Drug Design 6:585-606.
7. Eshed, Y. and Zamir, D. (1995). An introgression line population of Lycopersicon pennellii in the cultivated tomato enables the identification and fine mapping of yield-associated QTL. Genetics 141:1147-1162.
8. Fraser, P.D., Truesdale, M.R., Bird, C.R., Schuch, W., and Bramley, P.M. (1994). Carotenoid biosynthesis during tomato fruit development. Plant Physiol. 105:405-413.
9. Gillaspy, G., Ben-David, H., and Gruissem, W. (1993). Fruits - a developmental perspective. Plant Cell 5:1439-1451.
10. Grierson, D. and Schuch, W. (1993). Control of ripening. Philos.Trans.R.Soc.Lond.[Biol]. 342:241-250.
11. Hirschberg, J., Cohen, M., Harker, M., Lotan, T., Mann, V., and Pecker, I. (1997). Molecular genetics of the carotenoid biosynthesis pathway in plants and algae. Pure Appl.Chem. 69:2152-2158.
12. Laval-Martin, D., Quennement, J., and Moneger, R. (1975). Pigment evolution in Lycopersicon esculentum during growth and ripening. Biochemistry 14:2357-2362.
13. Mann, V., Pecker, I., and Hirschberg, J. (1994). Cloning and characterization of the gene for phytoene desaturase (*Pds*) from tomato (*Lycopersicon esculentum*). Plant Mol.Biol. 24:429-434.

14. Montgomery J, Goldman S, Deikman J, Margossian L, Fischer RL, (1993) Identification of an ethylene-responsive region in the promoter of a fruit-ripening gene. Proc. Natl. Acad. Sci. USA 90:5939-5943.
15. Nicholass FJ, Smith CJ, Schuch W, Bird CR, Grierson D. (1995) High levels of ripening-specific reporter gene expression directed by tomato fruit polygalacturonase gene-flanking regions. Plant Mol Biol 1995 28:423-435.
16. Montgomery J, Pollard V, Deikman J, Fischer RL. (1995) Positive and negative regulatory regions control the spatial distribution of polygalacturonase transcription in tomato fruit pericarp. Plant Cell 5:1049-1062
17. Pecker, I., Chamovitz, D., Linden, H., Sandmann, G., and Hirschberg, J. (1992). A single polypeptide catalyzing the conversion of phytoene to ζ -carotene is transcriptionally regulated during tomato fruit ripening. Proc.Natl.Acad.Sci.USA 89:4962-4966.
18. Pecker, I., Gabbay, R., Cunningham, F.X., and Hirschberg, J. (1996). Cloning and characterization of the cDNA for lycopene β-cyclase from tomato reveals decrease in its expression during fruit ripening. Plant Mol.Biol. 30:807-819.
19. Pnueli, L., Carmel-Goren, L., Hareven, D., Gutfinger, T., Alvarez, J., Ganal, M., Zamir, D., and Lifschitz, E. (1998). The SELF-PRUNING gene of tomato regulates vegetative to reproductive switching of sympodial meristems and is the ortholog of CEN and TFL1. Development 125:1979-1989.
20. Ray, J.A., Moureau, P., Bird, A.S., Grierson, D., Maunders, M., Truesdale, M., Bramley, P.M., and Schuch, W. (1992). Cloning and characterization of a gene involved in phytoene synthesis from tomato. Plant Mol.Biol. 19:401-404.
21. Sambrook, J., F, Fritch, E., F.,, and Maniatis, T., 1989. Molecular Cloning: A Laboratory Manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor. Sandmann, G. (1994a). Phytoene desaturase: Genes, enzymes and phylogenetic aspects. J.Plant Physiol. 143:444-447.
22. Sandmann, G. (1994b). Carotenoid biosynthesis in microorganisms and plants. Eur.J.Biochem. 223:7-24.
23. Tanksley, S.D., Ganal, M.W., Prince, J.C., de Vicente, M.C., Bonierabale, M.W., Broun, P., Fulton, T.M., Giovanonni, J.J., Grandillo, S., Martin, G.B., Messeguer, R., Miller, J.C., Miller, L., Paterson, A.H., Pineda, O., Roder, M.S., Wing, R.A., Wu, W., and Young, N.D. (1992).

High density molecular linkage maps of the tomato and potato genomes. Genetics 132:1141-1160.

24. Van Haaren MJ, Houck CM. (1991) Strong negative and positive regulatory elements contribute to the high-level fruit-specific expression of the tomato 2A11 gene. Plant Mol. Biol. 17:615-630

25. Wing, R.A., Zhang, H.B., and Tanksley, S.D. (1994). Map-based cloning in crop plants - tomato as a model system .1. genetic and physical mapping of jointless. Mol.Gen.Genet. 242:681-688.

26. Zhang, H.B., Martin, G.B., Tanksley, S.D., and Wing, R.A. (1994). Map-based cloning in crop plants: Tomato as a model system .2. Isolation and characterization of a set of overlapping yeast artificial chromosomes encompassing the jointless locus. Mol Gen Genet 244:613-621.

WHAT IS CLAIMED IS:

1. An isolated complementary or genomic DNA segment comprising a nucleotide sequence coding for a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, with the provision that said polypeptide has a major lycopene cyclase catalytic activity.

2. The isolated DNA segment of claim 1, wherein said nucleotide sequence is selected from the group consisting of SEQ ID NOs: 8, 9, 10 and 11 and functional naturally occurring and man-induced variants thereof.

3. The isolated DNA segment of claim 1, wherein said nucleotide sequence is a cDNA or a genomic DNA isolated form tomato.

4. An isolated complementary or genomic DNA segment comprising a nucleotide sequence selected from the group consisting of SEQ ID NOs: 8, 9, 10 and 11.

5. A polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, said polypeptide having a major lycopene cyclase catalytic activity.

6. A transduced cell overexpressing a polypeptide including an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, said polypeptide having a major lycopene cyclase catalytic activity, the cell therefore over producing β-carotene on an expense of lycopene.

7. The transduced cell of claim 6, selected from the group consisting of a prokaryotic cell and a eukaryotic cell.

8. The transduced cell of claim 7, wherein said eukaryotic cell is of a higher plant.

9. The transduced cell of claim 6, wherein the cell forms a part of a transgenic plant.

10. A method of down-regulating production of β-carotene in a cell comprising the step of introducing into the cell at least one anti-sense polynucleotide sequence capable of base pairing with messenger RNA coding for a polypeptide including an amino acid sequence selected from the group consisting of SEQ ID NOs: 17, 18 and 19 and functional naturally occurring and man-induced variants thereof, said polypeptide having a major lycopene cyclase catalytic activity, the cell therefore under producing β-carotene from lycopene.

11. The method of claim 10, wherein said at least one anti-sense polynucleotide sequence includes a synthetic oligonucleotide.

12. The method of claim 11, wherein said synthetic oligonucleotide includes a man-made modification rendering said synthetic oligonucleotide more stable in cell environment.

13. The method of claim 11, wherein said synthetic oligonucleotide is selected from the group consisting of methylphosphonate oligonucleotide, monothiophosphate oligonucleotide, dithiophosphate oligonucleotide, phosphoramidate oligonucleotide, phosphate ester oligonucleotide, bridged phosphorothioate oligonucleotide, bridged phosphoramidate oligonucleotide, bridged methylenephosphonate oligonucleotide, dephospho internucleotide analogs with siloxane bridges, carbonate bridge oligonucleotide, carboxymethyl ester bridge oligonucleotide, carbonate bridge oligonucleotide, carboxymethyl ester bridge oligonucleotide, acetamide bridge oligonucleotide, carbamate bridge oligonucleotide, thioether bridge oligonucleotide, sulfoxy bridge oligonucleotide, sulfono bridge oligonucleotide and α-anomeric bridge oligonucleotide.

14. The method of claim 10, wherein said at least one anti-sense polynucleotide sequence is encoded by an expression vector.

15. The method of claim 10, wherein said cell is selected from the group consisting of a prokaryotic cell and a eukaryotic cell.

16. The method of claim 15, wherein said eukaryotic cell is of a higher plant.

17. The method of claim 15, wherein the cell forms a part of a transgenic plant.

18. An expression construct for directing an expression of a gene in fruit or flower comprising a regulatory sequence selected from the group consisting of an upstream region of a B allele of tomato and an upstream region of a b allele of tomato.

19. The expression construct of claim 18, comprising a functional part of nucleotides 1-1210 of SEQ ID NO: 14 or nucleotides 1-1600 of SEQ ID NO: 15, or functional naturally occurring and man-induced variants thereof.

20. The expression construct of claim 18, comprising at least one control element having a sequence selected from the group consisting of SEQ ID NOs:21-24, all derived from SEQ ID NO:11, and functional naturally occurring and man-induced variants thereof.

21. The expression construct of claim 18, wherein the expression construct is selected from the group consisting of plasmid, cosmid, phage, virus, bacmid and artificial chromosome.

22. The expression construct of claim 18, designed to integrate into a genome of a host.

23. A method of isolating a gene encoding a polypeptide having an amino acid sequence homologous to SEQ ID NOs: 17, 18 and 19 and having a major lycopene cyclase catalytic activity from a species, the method comprising the step of screening a complementary or genomic DNA library prepared from isolated RNA or genomic DNA extracted from said species with a probe having a sequence derived from SEQ ID NOs: 8, 9, 10 or 11 and isolating clones reacting with said probe.

24. A transduced cell transduced with the expression construct of claim 18.

25. A transgenic plant transduced with the expression construct of claim 18.

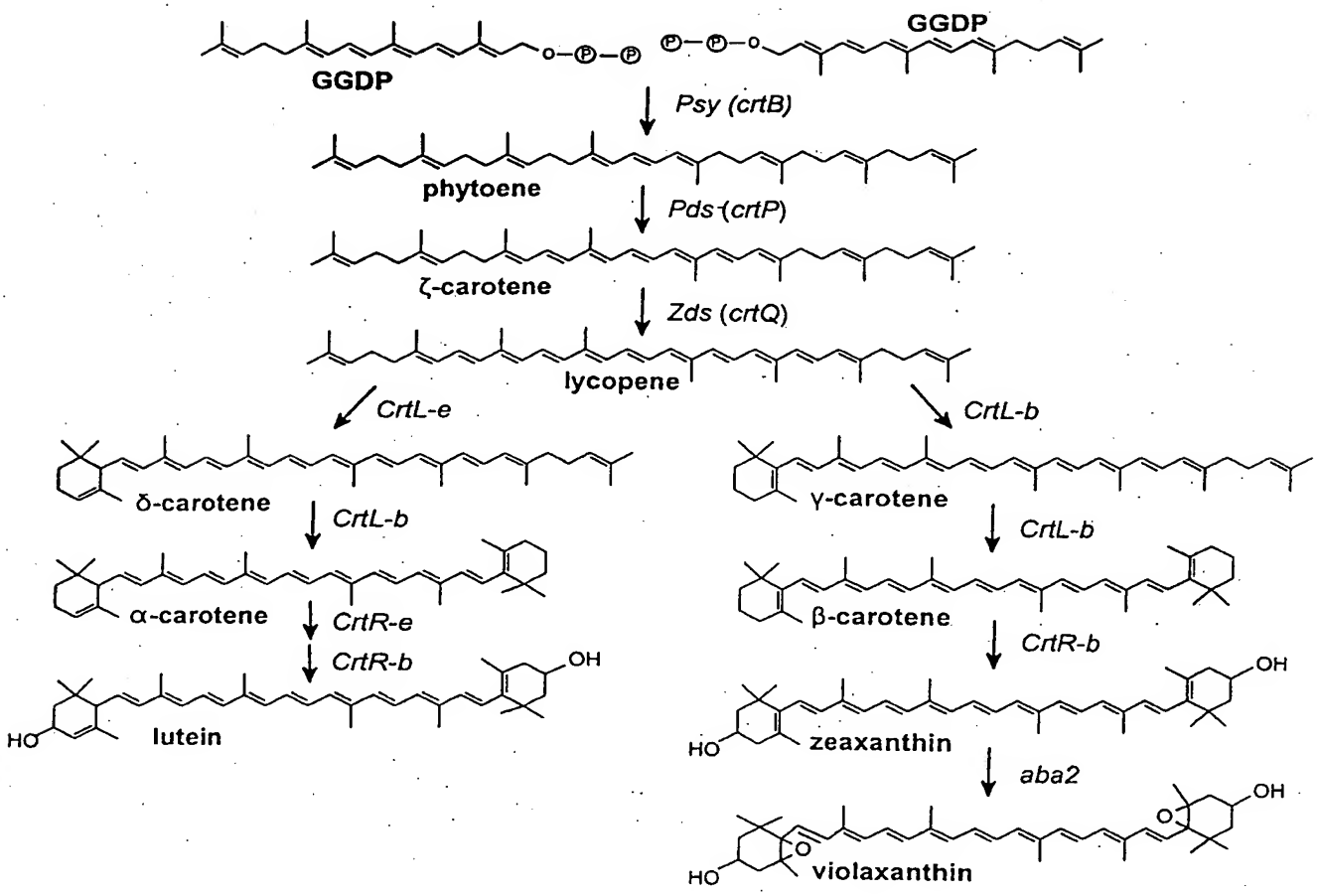
GGDP
GGDP
Psy (crtB)
phytoene
Pds (crtP)
ζ-carotene
Zds (crtQ)
lycopene
CrtL-e
CrtL-b
δ-carotene
γ-carotene
CrtL-b
CrtL-b
α-carotene
β-carotene
CrtR-e
CrtR-b
CrtR-b
OH
HO
lutein
zeaxanthin
aba2
violaxanthin
O

FIG. 1

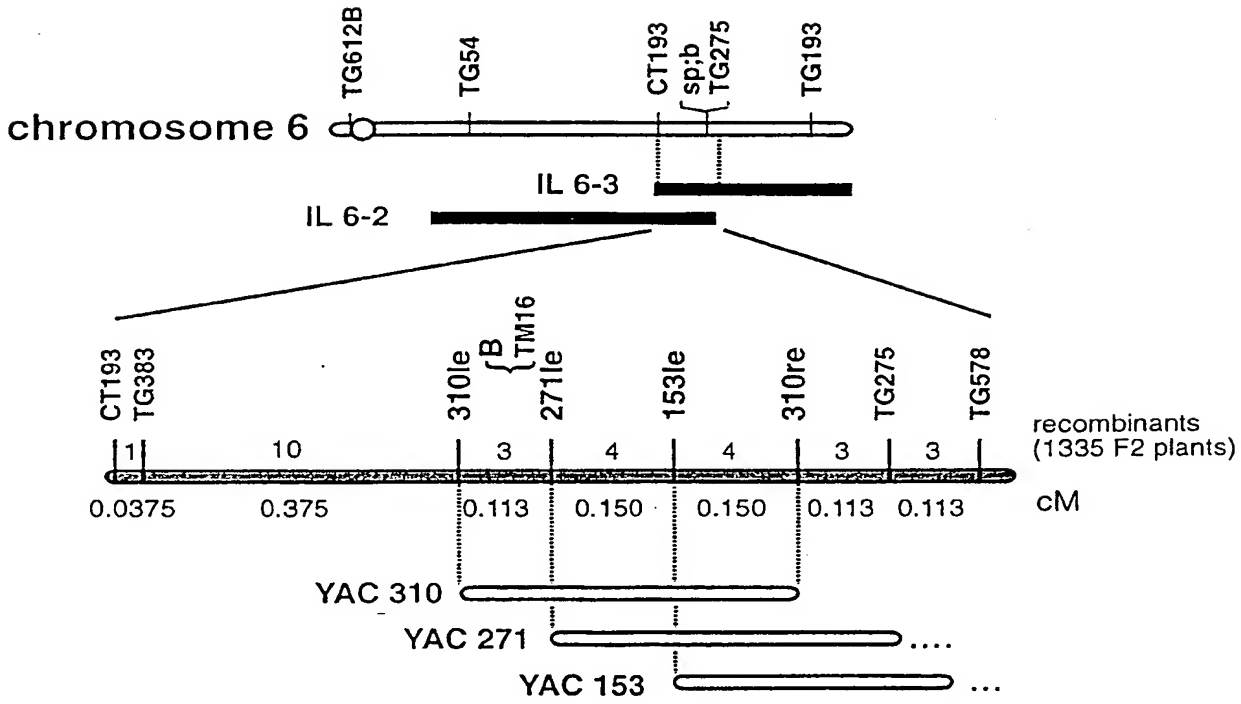
chromosome 6
TG612B
TG54
CT193
sp;b
TG275
TG193
IL 6-3
IL 6-2
CT193
TG383
310Ie
B
TM16
271Ie
153Ie
310re
TG275
TG578
recombinants
(1335 F2 plants)
1
10
3
4
4
3
3
0.0375
0.375
0.113
0.150
0.150
0.113
0.113
cM
YAC 310
YAC 271
YAC 153

FIG. 2

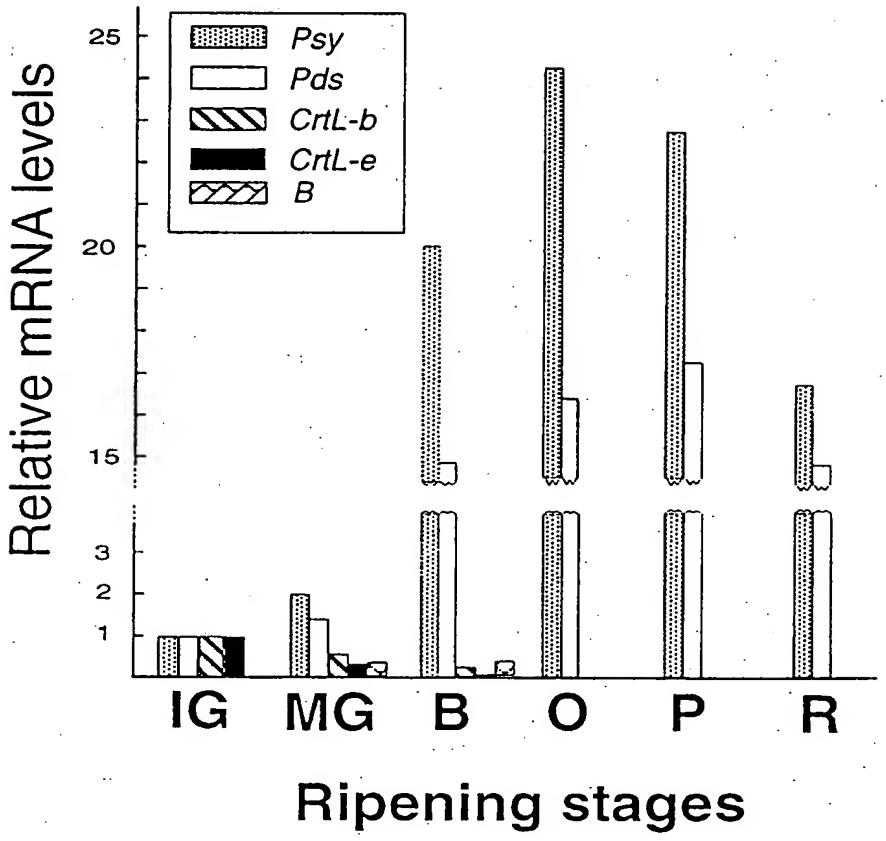
Relative mRNA levels
25
20
15
3
2
1
Psy
Pds
CrtL-b
CrtL-e
B
IG
MG
B
O
P
R
Ripening stages

FIG. 3

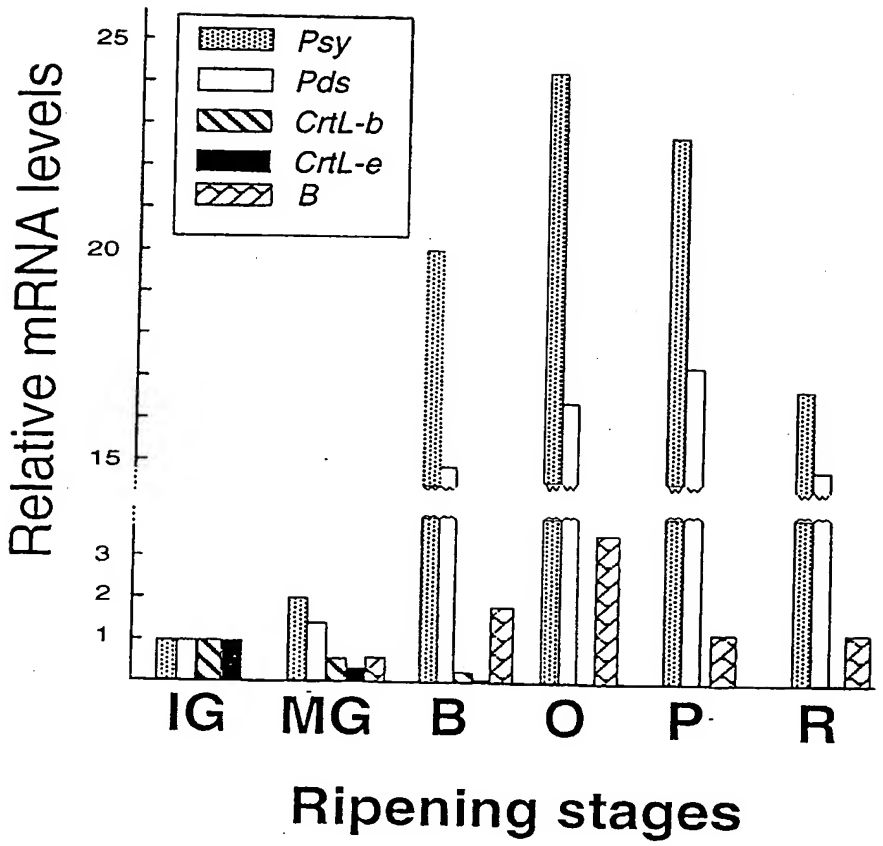
Relative mRNA levels
25
20
15
3
2
1
Psy
Pds
CrtL-b
CrtL-e
B
IG
MG
B
O
P
R
Ripening stages

FIG. 4

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Joseph Hirschberg et al.

(ii) TITLE OF INVENTION: POLYNUCLEOTIDES CONTROLLING THE EXPRESSION OF AND CODING FOR GENE B IN TOMATO AND USE OF SAME FOR ALTERING CAROTENOID BIOSYNTHESIS

(iii) NUMBER OF SEQUENCES: 25

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Mark M. Friedman c/o Anthony Castorina

(B) STREET: 20001 Jefferson Davis Highway, Suite 207

(C) CITY: Arlington

(D) STATE: Virginia

(E) COUNTRY: United States of America

(F) ZIP: 22202

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: 1.44 megabyte, 3.5" microdisk

(B) COMPUTER: Twinhead, Slimnote 890TX

(C) OPERATING SYSTEM: MS DOS version 6.2, Windows version 3.11

(D) SOFTWARE: Word for Windows version 2.0, converted to ASCI

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:

(B) FILING DATE:

(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER:

(B) FILING DATE:

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Friedmam, Mark M.

(B) REGISTRATION NUMBER: 33,883

(C) REFERENCE/DOCKET NUMBER: 325/12

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 972-3-5625553

(B) TELEFAX: 972-3-5625554

(C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 22

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

AATGGAAGCT CTTCTCAAGC CT 22

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

CACATTCAAA GGCTCTCTAT CGC 23

(2) INFORMATION FOR SEQ ID NO:3:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
TCGAGAACGG ACGATG 16

(2) INFORMATION FOR SEQ ID NO:4:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
TGCAGAGAGA CAGATG 16

(2) INFORMATION FOR SEQ ID NO:5:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
ATTTCATGCT TTATCTTTGA AG 22

(2) INFORMATION FOR SEQ ID NO:6:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
GCTGAAGTTG AAATTGTTGA 20

(2) INFORMATION FOR SEQ ID NO:7:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
TCTCTTCCTC AATAACACTT 20

(2) INFORMATION FOR SEQ ID NO:8:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1666
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

```
ATGGAAGCTC TTCTCAAGCC TTTTCCATCT CTTTTACTTT CCTCTCCTAC   50
ACCCCATAGG TCTATTTTCC AACAAAATCC CTCTTTTCTA AGTCCCACCA  100
CCAAAAAAAA ATCAAGAAAA TGTCTTCTTA GAAACAAAAG TAGTAAACTT  150
TTTTGTAGCT TTCTTGATTT AGCACCCACA TCAAAGCCAG AGTCTTTAGA  200
TGTTAACATC TCATGGGTTG ATCCTAATTC GAATCGGGCT CAATTCGACG  250
TGATCATTAT CGGAGCTGGC CCTGCTGGGC TCAGGCTAGC TGAACAAGTT  300
```

```
TCTAAATATG GTATTAAGGT ATGTTGTGTT GACCCTTCAC CACTCTCCAT  350
GTGGCCAAAT AATTATGGTG TTTGGGTTGA TGAGTTTGAG AATTTAGGAC  400
TGGAAAATTG TTTAGATCAT AAATGGCCTA TGACTTGTGT GCATATAAAT  450
GATAACAAAA CTAAGTATTT GGGAAGACCA TATGGTAGAG TTAGTAGAAA  500
GAAGCTGAAG TTGAAATTGT TGAATAGTTG TGTTGAGAAC AGAGTGAAGT  550
TTTATAAAGC TAAGGTTTGG AAAGTGGAAC ATGAAGAATT TGAGTCTTCA  600
ATTGTTTGTG ATGATGGTAA GAAGATAAGA GGTAGTTTGG TTGTGGATGC  650
AAGTGGTTTT GCTAGTGATT TTATAGAGTA TGACAGGCCA AGAAACCATG  700
GTTATCAAAT TGCTCATGGG GTTTTAGTAG AAGTTGATAA TCATCCATTT  750
GATTTGGATA AAATGGTGCT TATGGATTGG AGGGATTCTC ATTTGGGTAA  800
TGAGCCATAT TTAAGGGTGA ATAATGCTAA AGAACCAACA TTCTTGTATG  850
CAATGCCATT TGATAGAGAT TTGGTTTTCT TGGAAGAGAC TTCTTTGGTG  900
AGTCGTCCTG TTTTATCGTA TATGGAAGTA AAAAGAAGGA TGGTGGCAAG  950
ATTAAGGCAT TTGGGGATCA AAGTGAAAAG TGTTATTGAG GAAGAGAAAT 1000
GTGTGATCCC TATGGGAGGA CCACTTCCGC GGATTCCTCA AAATGTTATG 1050
GCTATTGGTG GGAATTCAGG GATAGTTCAT CCATCAACAG GGTACATGGT 1100
GGCTAGGAGC ATGGCTTTAG CACCAGTACT AGCTGAAGCC ATCGTCGAGG 1150
GGCTTGGCTC AACAAGAATG ATAAGAGGGT CTCAACTTTA CCATAGAGTT 1200
TGGAATGGTT TGTGGCCTTT GGATAGAAGA TGTGTTAGAG AATGTTATTC 1250
ATTTGGGATG GAGACATTGT TGAAGCTTGA TTTGAAAGGG ACTAGGAGAT 1300
TGTTTGACGC TTTCTTTGAT CTTGATCCTA AATACTGGCA AGGGTTCCTT 1350
TCTTCAAGAT TGTCTGTCAA AGAACTTGGT TTACTCAGCT TGTGTCTTTT 1400
CGGACATGGC TCAAACATGA CTAGGTTGGA TATTGTTACA AAATGTCCTC 1450
TTCCTTTGGT TAGACTGATT GGCAATCTAG CAATAGAGAG CCTTTGAATG 1500
TGAAAAGTTT GAATCATTTT CTTCATTTTA ATTTCTTTGA TTATTTTCAT 1550
ATTTTCTCAA TTGCAAAAGT GAGATAAGAG CTACATACTG TCAACAAATA 1600
AACTACTATT GGAAAGTTAA AATATGTGTT TGTTGTATGT TATTCTAATG 1650
GAATGGATTT TGTAAA                                      1666
```

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2876

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

```
GAATTCTCTG AAAAGGAGCA CCATATTTGC CGCACTGTGG TTCATATTTC   50
CAAGTACATT TAGATGAACT ATATCATCAG ATTGAAAGGT TATTGTATAA  100
TCAATCCAGT GGATTCTCGT TCTGGCACCT TTAGAAGTAC ATGTGCGGAA  150
AAGAATGATA AGGTTTGTAT TGTTGTTGAC AAAGCCTGTT GCCTTTCTCA  200
TTTGTAAATG TTCTGAACGA CTCCTAAATT ACTCTTAAGG TGTAAGGTCT  250
TCCGTGCCTG TTTGTAAATA TAATGCTGTG CCGTGACTTA CCTTTTGTAC  300
CATTTGTTCA AATGTATGGC CTGAACACCA GGGTTGTCAA AAATGTCTCA  350
TGCCCGTTTT ATTGGTCTGA AAATGGCGTG ATGCCAAATT CTGCCGCTCC  400
ACAGTGAGCA TTTCGATCTA CTGGAAATTG ACCAACTTAT TTTATCACTT  450
GATAACTAAA CAAAATCCTA TTAACTTTAA TCATACATTG TATTTATACC  500
GAAAAATTTA TGCATAACTC ATTAAATTAC CTTTTTTAGC AGTCAAATTC  550
TAAATCAGTT TCTAATTTAT CAAAATGGCT TTTATAGGGT CCCATTTCCA  600
CTAATATACC TGCCGTCCAT GCACTGACTA CAAAACAAAT ACCTCACTAT  650
GTTTGTTAGT GCTTGGTAAT ATAAAACCTT TTCTTTTATG AGAAAGTTCA  700
CCGAGAATAA TTTTCTATTT GTGGCATAAT AGTATATAGT GCAGATTGAC  750
AAGAATTTAA TTTTGCAGTT GGGCACATGA ACAATTTTCC TCAAAGTTGT  800
AGAAAGTACT TTTCATTTTC TTGTCACCGA AAATTATTTA TAATTGAAAT  850
TAAAACCGAA TGAGCTGCAA GATTCAAGTC GAATTTTCAA AAGAATTGAC  900
CAAGAAAAAA TTCAAAAATA TCCCCCACCC CCTACCAAAC ACATCCTAAA  950
GTGAGGTATA GACTGGGACT GGGATTGGGA AAAGGGTAAA ATGCTTTCAC 1000
TAGCTTAGCA AAGATTCCAC TTTGTTAGCT ATCTTTCTTT CTCATTTCCT 1050
TTTTTCTTTT TCTTTTTTTT GTTATATAAG CCAAAGTAGG TACCCAAAAG 1100
CATCAATATT TTGTATTGCT TGGTGATTCC TCTGTAGTCC AGTATTTCAT 1150
TTTCTACAAG TTCCACCTCC CTCCATAATT AACCATTATC AATCTTATAC 1200
ATTCTCTATA ATGGAAACTC TTCTCAAGCC TTTTCCATCT CTTTTACTTT 1250
CCTCTCCTAC ACCCCATAGG TCTATTTTCC AACAAAATCC CTCTTTTCTA 1300
AGTCCCACCA CCAAAAAAAA ATCAAGAAAA TGTCTTCTTA GAAACAAAAG 1350
TAGTAAACTT TTTTGTAGCT TTCTTGATTT AGCACCCACA TCAAAGCCAG 1400
AGTCTTTAGA TGTTAACATC TCATGGGTTG ATCCTAATTC GAATCGGGCT 1450
CAATTCGACG TGATCATTAT CGGAGCTGGC CCTGCTGGGC TCAGGCTAGC 1500
TGAACAAGTT TCTAAATATG GTATTAAGGT ATGTTGTGTT GACCCTTCAC 1550
CACTCTCCAT GTGGCCAAAT AATTATGGTG TTTGGGTTGA TGAGTTTGAG 1600
AATTTAGGAC TGGAAAATTG TTTAGATCAT AAATGGCCTA TGACTTGTGT 1650
GCATATAAAT GATAACAAAA CTAAGTATTT GGGAAGACCA TATGGTAGAG 1700
TTAGTAGAAA GAAGCTGAAG TTGAAATTGT TGAATAGTTG TGTTGAGAAC 1750
AGAGTGAAGT TTTATAAAGC TAAGGTTTGG AAAGTGGAAC ATGAAGAATT 1800
```

```
TGAGTCTTCA ATTGTTTGTG ATGATGGTAA GAAGATAAGA GGTAGTTTGG 1850
TTGTGGATGC AAGTGGTTTT GCTAGTGATT TTATAGAGTA TGACAGGCCA 1900
AGAAACCATG GTTATCAAAT TGCTCATGGG GTTTTAGTAG AAGTTGATAA 1950
TCATCCATTT GATTTGGATA AAATGGTGCT TATGGATTGG AGGGATTCTC 2000
ATTTGGGTAA TGAGCCATAT TTAAGGGTGA ATAATGCTAA AGAACCAACA 2050
TTCTTGTATG CAATGCCATT TGATAGAGAT TTGGTTTTCT TGGAAGAGAC 2100
TTCTTTGGTG AGTCGTCCTG TTTTATCGTA TATGGAAGTA AAAAGAAGGA 2150
TGGTGGCAAG ATTAAGGCAT TTGGGGATCA AAGTGAAAAG TGTTATTGAG 2200
GAAGAGAAAT GTGTGATCCC TATGGGAGGA CCACTTCCGC GGATTCCTCA 2250
AAATGTTATG GCTATTGGTG GGAATTCAGG GATAGTTCAT CCATCAACAG 2300
GGTACATGGT GGCTAGGAGC ATGGCTTTAG CACCAGTACT AGCTGAAGCC 2350
ATCGTCGAGG GGCTTGGCTC AACAAGAATG ATAAGAGGGT CTCAACTTTA 2400
CCATAGAGTT TGGAATGGTT TGTGGCCTTT GGATAGAAGA TGTGTTAGAG 2450
AATGTTATTC ATTTGGGATG GAGACATTGT TGAAGCTTGA TTTGAAAGGG 2500
ACTAGGAGAT TGTTTGACGC TTTCTTTGAT CTTGATCCTA AATACTGGCA 2550
AGGGTTCCTT TCTTCAAGAT TGTCTGTCAA AGAACTTGGT TTACTCAGCT 2600
TGTGTCTTTT CGGACATGGC TCAAACATGA CTAGGTTGGA TATTGTTACA 2650
AAATGTCCTC TTCCTTTGGT TAGACTGATT GGCAATCTAG CAATAGAGAG 2700
CCTTTGAATG TGAAAAGTTT GAATCATTTT CTTCATTTTA ATTTCTTTGA 2750
TTATTTTCAT ATTTTCTCAA TTGCAAAAGT GAGATAAGAG CTACATACTG 2800
TCAACAAATA AACTACTATT GGAAAGTTAA AATATGTGTT TGTTGTATGT 2850
TATTCTAATG GAATGGATTT TGTAAA                           2876
```

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1740

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```
ATGGAAGCTC TTCTCAAGCC TTTTCCATCT CTTTTACTTT CCTCTCCTAC   50
ACCCTATAGG TCTATTGTCC AACAAAATCC TTCTTTTCTA AGTCCCACCA  100
CCAAAAAAAA TCAAGAAAAT GTCTTCTTAG AAACAAAAGT AGTAAACTTT  150
TTTGTAGCTT TCTTGATTTA GCACCCACAT CAAAGCCAGA GTCTTTAAAT  200
GTTAACATCT CATGGGTTGA TCCTAATTCG AATCGGGCTC AATTCGACGT  250
GATCATTATC GGAGCTGGCC CTGCTGGGCT CAGGCTAGCT GAACAAGTTT  300
CTAAATATGG TATTAAGGTA TGTTGTGTTG ACCCTTCACC ACTCTCCATG  350
TGGCCAAATA ATTATGGTGT TTGGGTTGAT GAGTTTGAGA ATTTAGGACT  400
GGAAAATTGT TTAGATCATA AATGGCCTAT GACTTGTGTG CATATAAATG  450
ATAACAAAAC TAAGTATTTG GGAAGACCAT ATGGTAGAGT TAGTAGAAAG  500
AAGCTGAAGT TGAAATTGTT GAATAGTTGT GTTGAGAACA GAGTGAAGTT  550
TTATAAAGCT AAGGTTTGGA AAGTGGAACA TGAAGAATTT GAGTCTTCAA  600
TTGTTTGTGA TGATGGTAAG AAGATAAGAG GTAGTTTGGT TGTGGATGCA  650
AGTGGTTTTG CTAGTGATTT TATAGAGTAT GACAGGCCAA GAAACCATGG  700
TTATCAAATT GCTCATGGGG TTTTAGTAGA AGTTGATAAT CATCCATTTG  750
ATTTGGATAA AATGGTGCTT ATGGATTGGA GGGATTCTCA TTTGGGTAAT  800
GAGCCATATT TAAGGGTGAA TAATGCTAAA GAACCAACAT TCTTGTATGC  850
AATGCCATTT GATAGAGATT TGGTTTTCTT GGAAGAGACT TCTTTGGTGA  900
GTCGTCCTGT GTTATCGTAT ATGGAAGTAA AAAGAAGGAT GGTGGCAAGA  950
TTAAGGCATT TGGGGATCAA AGTGAAAAGT GTTATTGAGG AAGAGAAATG 1000
TGTGATCCCT ATGGGAGGAC CACTTCCGCG GATTCCTCAA AATGTTATGG 1050
CTATTGGTGG GAATTCAGGG ATAGTTCATC CATCAACAGG GTACATGGTG 1100
GCTAGGAGCA TGGCTTTAGC ACCAGTACTA GCTGAAGCCA TCGTCGAGGG 1150
GCTTGGCTCA ACAAGAATGA TAAGAGGGTC TCAACTTTAC CATAGAGTTT 1200
GGAATGGTTT GTGGCCTTTG GATAGAAGAT GTGTTAGAGA ATGTTATTCA 1250
TTTGGGATGG AGACATTGTT GAAGCTTGAT TTGAAAGGGA CTAGGAGATT 1300
GTTTGACGCT TTCTTTGATC TTGATCCTAA ATACTGGCAA GGGTTCCTTT 1350
CTTCAAGATT GTCTGTCAAA GAAACTTGGT TTACTCAGCT TGTGTCTTTT 1400
CGGACATGGC TCAAACATGA CTAGGTTGGG ATATTGTTAC AAAATGTCCT 1450
CTTCCTTTGG TTAGACTGAT TGGCAATCTA GCAATAGAGA GCCTTTGAAA 1500
TGTGAAAAGT TTGAATCATT TTCTTCATTT TAATTTCTTT GATTATTTTC 1550
ATATTTTCTC AATTGCAGAA TGAGATAAAA ACTACATACT GTCGACAAAT 1600
AAACTACTAT TGGAANGTTA AAATAATGTG TGTGTTGNAT GTTANGCCTA 1650
ATGGAANGGA TGNGGTTANG CAATTTATGA ACTGNNCGCT CTGTTCGCTT 1700
AAAANCCTTG GTTCCACCTT AANGGAANGG NCCGGCCATT           1740
```

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2897

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TGGTTCATAT TTCCAATTAC ATTTAGATGA ACTATATCAT CAGGAGTGAA 50
AGGTTATTGT ATAATCAATC CAGTGGATTC TCGTTCTGGC ACCTTTAGAA 100
GTACATGTGC GGAAAAGAAT GATAAGGTTT GTATTGTTGT TGACAAGGCC 150
TGTTGCCTTT CTCATTTGTA AATGTTCTGA ACGACTCCTA AATTACTCTT 200
AAAGTGTAAG GTCTTCCGTG CCTGTTTGTA TATATAATGC TGTGCCGTGA 250
CTTACCTTTT GTACCATTTG TTCAAATGTA TGGCCTGGAC ACTAGGGTTG 300
TCAAAAATGT CTCATGACTT CACCCTTCTT TCTTGTCTTG GTGCCCGTTT 350
TATTGGTCTG AGAACGGCGT GATGCCAAAT TCTGCCGCTC CACAGTGAGC 400
ATTTCGATCT ACTGGAAATT GACCAACTTA TTTTATCACT TGATAACTAG 450
AGTCTGGGTT CAAACAAAAT CCAATAACTT CAATCATACA TTGTATTTAT 500
ATTGAAAAAA TTATGCACAA CTCAGTAAAT TACCTTTTTT TGCAGTCAAA 550
AATTCTAGAT CAGTTTCTAA TTAATCAAAA TGGCCTTTAT AGGGTCCCAG 600
TTCCATTAAT ATACCTGCCG TCCATGCACT GATTACAAGA CAAATACCTC 650
ACTATGTTTG TTAGTGCTTG GTAATATAAA ACCTTTTCTT TTATGAGAAA 700
GTTCACCGAA AATAATTTTC TATTTGTGGC ATAACTAGTA TCGAAGTATA 750
TAGTGCAGAT TGACAAGAAT TTAATTTTGC AGTTGGGCAC ATGAACAATT 800
TTCCTCAAAG TTGTAGAAAA TATTTTTCAT TTTCTTGTCA CCGAAAATTA 850
TTTATAATTG AAATTGAAAC CGAATGAGCT GCAAGACTCG AGTCGAATTT 900
CAAAAAAATT GACCAACTAA ATATGAAAAA ATCCGAATAT ATCCCCCACC 950
CCCTACCAAA CACATCCTAA AGTGAGGTAT AGACTGGGAC TGGGATTGGG 1000
AAAAGGGTAA AATGCTTTCA CTAGCTTAGC AAAGATTCCA CTTTGTTAGC 1050
TATCTTTCTT TCTCATTTCC TTTTTTCTTT TTCTTTTTTT TGTTATATAA 1100
GCCAAAGTAG GTACCCAAAA GCATCAATAT TTTGTATTGC TTGGTGATTC 1150
CTCTTTACTC CAGTATTTCA TTTTCTACAA GTTCCACCTC CCTCCATAAT 1200
TAACCATTAT CAATCTTATA CATTTTCTAT AATGGAAACT CTTCTCAAGC 1250
CTTTTCCATC TCTTTTACTT TCCTCTCCTA CACCCTATAG GTCTATTGTC 1300
CAACAAAATC CTTCTTTTCT AAGTCCCACC ACCCAAAAAA AATCAAGAAA 1350
ATGTCTTCTT AGAAACAAAA GTAGTAAACT TTTTTGTAGC TTTCTTGATT 1400
TAGCACCCAC ATCAAAGCCA GAGTCTTTAA ATGTTAACAT CTCATGGGTT 1450
GATCCTAATT CTGGTCGGGC TCAATTCGAC GTGATCATTA TCGGAGCTGG 1500
CCCTGCTGGG CTCAGGTTAG CTGAACAAGT TTCTAAATAT GGTATTAAGG 1550
TATGTTGTGT TGACCCTTCA CCACTCTCCA TGTGGCCAAA TAATTATGGT 1600
GTTTGGGTTG ATGAGTTTGA GAATTTAGGA CTGGAAGATT GTTTAGATCA 1650
TAAATGGCCT ATGACTTGTG TGCATATAAA TGATAACAAG ACTAAGTATT 1700
TGGGAAGACC ATATGGTAGA GTTAGTAGAA AGAAGCTGAA GTTGAAATTG 1750
TTGAACAGTT GTGTTGAGAA CAGAGTGAAG TTTTATAAAG CTAAGGTTTG 1800
GAAAGTGGAA CATGAAGAAT TTGAGTCTTC AATTGTTTGT GATGATGGTA 1850
AGAAGATAAG AGGTAGTTTG GTTGTGGATG CAAGTGGTTT TGCTAGTGAT 1900
TTTATAGAGT ATGACAAGCC AAGAAACCAT GGTTATCAAA TTGCTCATGG 1950
GGTTTTAGTA GAAGTTGATA ATCATCCATT TGATTTGGAT AAAATGGTGC 2000
TTATGGATTG GAGGGATTCT CATTTAGGTA ATGAGCCATA TTTAAGGGTG 2050
AATAATGCTA AAGAACCAAC ATTCTTGTAT GCAATGCCAT TTGATAGAAA 2100
TTTGGTTTTC TTGGAAGAGA CTTCTTTGGT GAGTCGTCCT GTGTTATCGT 2150
ATATGGAAGT AAAAAGAAGG ATGGTGGCAA GATTAAGGCA TTTGGGGATC 2200
AAAGTGAGAA GTGTTATTGA GGAAGAGAAA TGTGTGATCC CTATGGGAGG 2250
ACCACTTCCG CGGATTCCTC AAAATGTTAT GGCTATTGGT GGGAATTCAG 2300
GGATAGTTCA TCCATCAACG GGGTACATGG TGGCTAGGAG CATGGCTTTA 2350
GCACCAGTAC TAGCTGAAGC CATCGTCGAG GGGCTTGGCT CAACAAGAAT 2400
GATAAGAGGG TCTCAACTTT ACCATAGAGT TTGGAATGGT TTGTGGCCTT 2450
TGGATAGAAG ATGTGTTAGA GAATGTTATT CATTTGGGAT GGAGACATTG 2500
TTGAAGCTTG ATTTGAAAGG GACTAGGAGA TTGTTTGACG CTTTCTTTGA 2550
TCTTGATCCT AAATACTGGC AAGGGTTCCT TTCTTCAAGA TTGTCTGTCA 2600
AAGAACTTGG TTTACTCAGC TTGTGTCTTT TCGGACATGG CTCAAATTTG 2650
ACTAGGTTGG ATATTGTTAC AAAATGTCCT GTTCCTTTGG TTAGACTGAT 2700
TGGCAATCTA GCAGTAGAGA GCCTTTGAAT GTGAAAAGTT TGAATCATTT 2750
TCTTTATTTT AATTTCTTTG ATTATTTTCA TATTTTCTCA ATGCAAAAGT 2800
GAGAGAAGAC TATACACTGT CAACAAATAA ACTACTATTG GAAAGTTAAA 2850
ATAATGTGTG TGTTGTATGT TATGCTAATG GAATGGATTG GTGTAAA 2897

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1740
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

ATGGAAGCTC TTCTCAAGCC TTTTCCATCT CTTTTACTTT CCTCTCCTAC 50
ACCCTATAGG TCTATTGTCC AACAAAATCC TTCTTTTCTA AGTCCCACCA 100

```
CCAAAAAAAA TCAAGAAAAT GTCTTCTTAG AAACAAAAGT AGTAAACTTT  150
TTTGTAGCTT TCTTGATTTA GCACCCACAT CAAAGCCAGA GTCTTTAAAT  200
GTTAACATCT CATGGGTTGA TCCTAATTCG AATCGGGCTC AATTCGACGT  250
GATCATTATC GGAGCTGGCC CTGCTGGGCT CAGGCTAGCT GAACAAGTTT  300
CTAAATATGG TATTAAGGTA TGTTGTGTTG ACCCTTCACC ACTCTCCATG  350
TGGCCAAATA ATTATGGTGT TTGGGTTGAT GAGTTTGAGA ATTTAGGACT  400
GGAAAATTGT TTAGATCATA AATGGCCTAT GACTTGTGTG CATATAAATG  450
ATAACAAAAC TAAGTATTTG GGAAGACCAT ATGGTAGAGT TAGTAGAAAG  500
AAGCTGAAGT TGAAATTGTT GAATAGTTGT GTTGAGAACA GAGTGAAGTT  550
TTATAAAGCT AAGGTTTGGA AAGTGGAACA TGAAGAATTT GAGTCTTCAA  600
TTGTTTGTGA TGATGGTAAG AAGATAAGAG GTAGTTTGGT TGTGGATGCA  650
AGTGGTTTTG CTAGTGATTT TATAGAGTAT GACAGGCCAA GAAACCATGG  700
TTATCAAATT GCTCATGGGG TTTTAGTAGA AGTTGATAAT CATCCATTTG  750
ATTTGGATAA AATGGTGCTT ATGGATTGGA GGGATTCTCA TTTGGGTAAT  800
GAGCCATATT TAAGGGTGAA TAATGCTAAA GAACCAACAT TCTTGTATGC  850
AATGCCATTT GATAGAGATT TGGTTTTCTT GGAAGAGACT TCTTTGGTGA  900
GTCGTCCTGT GTTATCGTAT ATGGAAGTAA AAAGAAGGAT GGTGGCAAGA  950
TTAAGGCATT TGGGGATCAA AGTGAAAAGT GTTATTGAGG AAGAGAAATG 1000
TGTGATCCCT ATGGGAGGAC CACTTCCGCG GATTCCTCAA AATGTTATGG 1050
CTATTGGTGG GAATTCAGGG ATAGTTCATC CATCAACAGG GTACATGGTG 1100
GCTAGGAGCA TGGCTTTAGC ACCAGTACTA GCTGAAGCCA TCGTCGAGGG 1150
GCTTGGCTCA ACAAGAATGA TAAGAGGGTC TCAACTTTAC CATAGAGTTT 1200
GGAATGGTTT GTGGCCTTTG GATAGAAGAT GTGTTAGAGA ATGTTATTCA 1250
TTTGGGATGG AGACATTGTT GAAGCTTGAT TTGAAAGGGA CTAGGAGATT 1300
GTTTGACGCT TTCTTTGATC TTGATCCTAA ATACTGGCAA GGGTTCCTTT 1350
CTTCAAGATT GTCTGTCAAA GAAACTTGGT TTACTCAGCT TGTGTCTTTT 1400
CGGACATGGC TCAAACATGA CTAGGTTGGG ATATTGTTAC AAAATGTCCT 1450
CTTCCTTTGG TTAGACTGAT TGGCAATCTA GCAATAGAGA GCCTTTGAAA 1500
TGTGAAAAGT TTGAATCATT TTCTTCATTT TAATTTCTTT GATTATTTTC 1550
ATATTTTCTC AATTGCAGAA TGAGATAAAA ACTACATACT GTCGACAAAT 1600
AAACTACTAT TGGAANGTTA AAATAATGTG TGTGTTGNAT GTTANGCCTA 1650
ATGGAANGGA TGNGGTTANG CAATTTATGA ACTGNNCGCT CTGTTCGCTT 1700
AAAANCCTTG GTTCCACCTT AANGGAANGG NCCGGCCATT             1740
```

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1666

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

```
ATG GAA GCT CTT CTC AAG CCT TTT CCA TCT CTT TTA CTT TCC TCT   45
Met Glu Ala Leu Leu Lys Pro Phe Pro Ser Leu Leu Leu Ser Ser
  5                  10                  15
CCT ACA CCC CAT AGG TCT ATT TTC CAA CAA AAT CCC TCT TTT CTA   90
Pro Thr Pro His Arg Ser Ile Phe Gln Gln Asn Pro Ser Phe Leu
             20                  25                  30
AGT CCC ACC ACC AAA AAA AAA TCA AGA AAA TGT CTT CTT AGA AAC  135
Ser Pro Thr Thr Lys Lys Lys Ser Arg Lys Cys Leu Leu Arg Asn
         35                  40                  45
AAA AGT AGT AAA CTT TTT TGT AGC TTT CTT GAT TTA GCA CCC ACA  180
Lys Ser Ser Lys Leu Phe Cys Ser Phe Leu Asp Leu Ala Pro Thr
     50                  55                  60
TCA AAG CCA GAG TCT TTA GAT GTT AAC ATC TCA TGG GTT GAT CCT  225
Ser Lys Pro Glu Ser Leu Asp Val Asn Ile Ser Trp Val Asp Pro
 65                  70                  75
AAT TCG AAT CGG GCT CAA TTC GAC GTG ATC ATT ATC GGA GCT GGC  270
Asn Ser Asn Arg Ala Gln Phe Asp Val Ile Ile Ile Gly Ala Gly
             80                  85                  90
CCT GCT GGG CTC AGG CTA GCT GAA CAA GTT TCT AAA TAT GGT ATT  315
Pro Ala Gly Leu Arg Leu Ala Glu Gln Val Ser Lys Tyr Gly Ile
         95                 100                 105
AAG GTA TGT TGT GTT GAC CCT TCA CCA CTC TCC ATG TGG CCA AAT  360
Lys Val Cys Cys Val Asp Pro Ser Pro Leu Ser Met Trp Pro Asn
        110                 115                 120
AAT TAT GGT GTT TGG GTT GAT GAG TTT GAG AAT TTA GGA CTG GAA  405
Asn Tyr Gly Val Trp Val Asp Glu Phe Glu Asn Leu Gly Leu Glu
    125                 130                 135
AAT TGT TTA GAT CAT AAA TGG CCT ATG ACT TGT GTG CAT ATA AAT  450
Asn Cys Leu Asp His Lys Trp Pro Met Thr Cys Val His Ile Asn
140                 145                 150
GAT AAC AAA ACT AAG TAT TTG GGA AGA CCA TAT GGT AGA GTT AGT  495
```

```
Asp Asn Lys Thr Lys Tyr Leu Gly Arg Pro Tyr Gly Arg Val Ser
                155                 160                 165
AGA AAG AAG CTG AAG TTG AAA TTG TTG AAT AGT TGT GTT GAG AAC  540
Arg Lys Lys Leu Lys Leu Lys Leu Leu Asn Ser Cys Val Glu Asn
                170                 175                 180
AGA GTG AAG TTT TAT AAA GCT AAG GTT TGG AAA GTG GAA CAT GAA  585
Arg Val Lys Phe Tyr Lys Ala Lys Val Trp Lys Val Glu His Glu
                185                 190                 195
GAA TTT GAG TCT TCA ATT GTT TGT GAT GAT GGT AAG AAG ATA AGA  630
Glu Phe Glu Ser Ser Ile Val Cys Asp Asp Gly Lys Lys Ile Arg
                200                 205                 210
GGT AGT TTG GTT GTG GAT GCA AGT GGT TTT GCT AGT GAT TTT ATA  675
Gly Ser Leu Val Val Asp Ala Ser Gly Phe Ala Ser Asp Phe Ile
                215                 220                 225
GAG TAT GAC AGG CCA AGA AAC CAT GGT TAT CAA ATT GCT CAT GGG  720
Glu Tyr Asp Arg Pro Arg Asn His Gly Tyr Gln Ile Ala His Gly
                230                 235                 240
GTT TTA GTA GAA GTT GAT AAT CAT CCA TTT GAT TTG GAT AAA ATG  765
Val Leu Val Glu Val Asp Asn His Pro Phe Asp Leu Asp Lys Met
                245                 250                 255
GTG CTT ATG GAT TGG AGG GAT TCT CAT TTG GGT AAT GAG CCA TAT  810
Val Leu Met Asp Trp Arg Asp Ser His Leu Gly Asn Glu Pro Tyr
                260                 265                 270
TTA AGG GTG AAT AAT GCT AAA GAA CCA ACA TTC TTG TAT GCA ATG  855
Leu Arg Val Asn Asn Ala Lys Glu Pro Thr Phe Leu Tyr Ala Met
                275                 280                 285
CCA TTT GAT AGA GAT TTG GTT TTC TTG GAA GAG ACT TCT TTG GTG  900
Pro Phe Asp Arg Asp Leu Val Phe Leu Glu Glu Thr Ser Leu Val
                290                 295                 300
AGT CGT CCT GTT TTA TCG TAT ATG GAA GTA AAA AGA AGG ATG GTG  945
Ser Arg Pro Val Leu Ser Tyr Met Glu Val Lys Arg Arg Met Val
                305                 310                 315
GCA AGA TTA AGG CAT TTG GGG ATC AAA GTG AAA AGT GTT ATT GAG  990
Ala Arg Leu Arg His Leu Gly Ile Lys Val Lys Ser Val Ile Glu
                320                 325                 330
GAA GAG AAA TGT GTG ATC CCT ATG GGA GGA CCA CTT CCG CGG ATT 1035
Glu Glu Lys Cys Val Ile Pro Met Gly Gly Pro Leu Pro Arg Ile
                335                 340                 345
CCT CAA AAT GTT ATG GCT ATT GGT GGG AAT TCA GGG ATA GTT CAT 1080
Pro Gln Asn Val Met Ala Ile Gly Gly Asn Ser Gly Ile Val His
                350                 355                 360
CCA TCA ACA GGG TAC ATG GTG GCT AGG AGC ATG GCT TTA GCA CCA 1125
Pro Ser Thr Gly Tyr Met Val Ala Arg Ser Met Ala Leu Ala Pro
                365                 370                 375
GTA CTA GCT GAA GCC ATC GTC GAG GGG CTT GGC TCA ACA AGA ATG 1170
Val Leu Ala Glu Ala Ile Val Glu Gly Leu Gly Ser Thr Arg Met
                380                 385                 390
ATA AGA GGG TCT CAA CTT TAC CAT AGA GTT TGG AAT GGT TTG TGG 1215
Ile Arg Gly Ser Gln Leu Tyr His Arg Val Trp Asn Gly Leu Trp
                395                 400                 405
CCT TTG GAT AGA AGA TGT GTT AGA GAA TGT TAT TCA TTT GGG ATG 1260
Pro Leu Asp Arg Arg Cys Val Arg Glu Cys Tyr Ser Phe Gly Met
                410                 415                 420
GAG ACA TTG TTG AAG CTT GAT TTG AAA GGG ACT AGG AGA TTG TTT 1305
Glu Thr Leu Leu Lys Leu Asp Leu Lys Gly Thr Arg Arg Leu Phe
                425                 430                 435
GAC GCT TTC TTT GAT CTT GAT CCT AAA TAC TGG CAA GGG TTC CTT 1350
Asp Ala Phe Phe Asp Leu Asp Pro Lys Tyr Trp Gln Gly Phe Leu
                440                 445                 450
TCT TCA AGA TTG TCT GTC AAA GAA CTT GGT TTA CTC AGC TTG TGT 1395
Ser Ser Arg Leu Ser Val Lys Glu Leu Gly Leu Leu Ser Leu Cys
                455                 460                 465
CTT TTC GGA CAT GGC TCA AAC ATG ACT AGG TTG GAT ATT GTT ACA 1440
Leu Phe Gly His Gly Ser Asn Met Thr Arg Leu Asp Ile Val Thr
                470                 475                 480
AAA TGT CCT CTT CCT TTG GTT AGA CTG ATT GGC AAT CTA GCA ATA 1485
Lys Cys Pro Leu Pro Leu Val Arg Leu Ile Gly Asn Leu Ala Ile
                485                 490                 495
GAG AGC CTT TGA ATG TGA AAA GTT TGA ATC ATT TTC TTC ATT TTA 1530
Glu Ser Leu
        498
ATT TCT TTG ATT ATT TTC ATA TTT TCT CAA TTG CAA AAG TGA GAT 1575
AAG AGC TAC ATA CTG TCA ACA AAT AAA CTA CTA TTG GAA AGT TAA 1620
AAT ATG TGT TTG TTG TAT GTT ATT CTA ATG GAA TGG ATT TTG TAA 1665
A                                                          1666
```

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2876

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

```
  G AAT TCT CTG AAA AGG AGC ACC ATA TTT GCC GCA CTG TGG TTC   43
ATA TTT CCA AGT ACA TTT AGA TGA ACT ATA TCA TCA GAT TGA AAG   88
GTT ATT GTA TAA TCA ATC CAG TGG ATT CTC GTT CTG GCA CCT TTA  133
GAA GTA CAT GTG CGG AAA AGA ATG ATA AGG TTT GTA TTG TTG TTG  178
ACA AAG CCT GTT GCC TTT CTC ATT TGT AAA TGT TCT GAA CGA CTC  223
CTA AAT TAC TCT TAA GGT GTA AGG TCT TCC GTG CCT GTT TGT AAA  268
TAT AAT GCT GTG CCG TGA CTT ACC TTT TGT ACC ATT TGT TCA AAT  313
GTA TGG CCT GAA CAC CAG GGT TGT CAA AAA TGT CTC ATG CCC GTT  358
TTA TTG GTC TGA AAA TGG CGT GAT GCC AAA TTC TGC CGC TCC ACA  403
GTG AGC ATT TCG ATC TAC TGG AAA TTG ACC AAC TTA TTT TAT CAC  448
TTG ATA ACT AAA CAA AAT CCT ATT AAC TTT AAT CAT ACA TTG TAT  493
TTA TAC CGA AAA ATT TAT GCA TAA CTC ATT AAA TTA CCT TTT TTA  538
GCA GTC AAA TTC TAA ATC AGT TTC TAA TTT ATC AAA ATG GCT TTT  583
ATA GGG TCC CAT TTC CAC TAA TAT ACC TGC CGT CCA TGC ACT GAC  628
TAC AAA ACA AAT ACC TCA CTA TGT TTG TTA GTG CTT GGT AAT ATA  673
AAA CCT TTT CTT TTA TGA GAA AGT TCA CCG AGA ATA ATT TTC TAT  718
TTG TGG CAT AAT AGT ATA TAG TGC AGA TTG ACA AGA ATT TAA TTT  763
TGC AGT TGG GCA CAT GAA CAA TTT TCC TCA AAG TTG TAG AAA GTA  808
CTT TTC ATT TTC TTG TCA CCG AAA ATT ATT TAT AAT TGA AAT TAA  853
AAC CGA ATG AGC TGC AAG ATT CAA GTC GAA TTT TCA AAA GAA TTG  898
ACC AAG AAA AAA TTC AAA AAT ATC CCC CAC CCC CTA CCA AAC ACA  943
TCC TAA AGT GAG GTA TAG ACT GGG ACT GGG ATT GGG AAA AGG GTA  988
AAA TGC TTT CAC TAG CTT AGC AAA GAT TCC ACT TTG TTA GCT ATC 1033
TTT CTT TCT CAT TTC CTT TTT TCT TTT TCT TTT TTT TGT TAT ATA 1078
AGC CAA AGT AGG TAC CCA AAA GCA TCA ATA TTT TGT ATT GCT TGG 1123
TGA TTC CTC TGT AGT CCA GTA TTT CAT TTT CTA CAA GTT CCA CCT 1168
CCC TCC ATA ATT AAC CAT TAT CAA TCT TAT ACA TTC TCT ATA ATG 1213
                                                       Met
GAA ACT CTT CTC AAG CCT TTT CCA TCT CTT TTA CTT TCC TCT CCT 1258
Glu Thr Leu Leu Lys Pro Phe Pro Ser Leu Leu Leu Ser Ser Pro
                5                  10                  15
ACA CCC CAT AGG TCT ATT TTC CAA CAA AAT CCC TCT TTT CTA AGT 1303
Thr Pro His Arg Ser Ile Phe Gln Gln Asn Pro Ser Phe Leu Ser
            20                  25                  30
CCC ACC ACC AAA AAA AAA TCA AGA AAA TGT CTT CTT AGA AAC AAA 1348
Pro Thr Thr Lys Lys Lys Ser Arg Lys Cys Leu Leu Arg Asn Lys
        35                  40                  45
AGT AGT AAA CTT TTT TGT AGC TTT CTT GAT TTA GCA CCC ACA TCA 1393
Ser Ser Lys Leu Phe Cys Ser Phe Leu Asp Leu Ala Pro Thr Ser
    50                  55                  60
AAG CCA GAG TCT TTA GAT GTT AAC ATC TCA TGG GTT GAT CCT AAT 1438
Lys Pro Glu Ser Leu Asp Val Asn Ile Ser Trp Val Asp Pro Asn
65                  70                  75
TCG AAT CGG GCT CAA TTC GAC GTG ATC ATT ATC GGA GCT GGC CCT 1483
Ser Asn Arg Ala Gln Phe Asp Val Ile Ile Ile Gly Ala Gly Pro
            80                  85                  90
GCT GGG CTC AGG CTA GCT GAA CAA GTT TCT AAA TAT GGT ATT AAG 1528
Ala Gly Leu Arg Leu Ala Glu Gln Val Ser Lys Tyr Gly Ile Lys
        95                  100                 105
GTA TGT TGT GTT GAC CCT TCA CCA CTC TCC ATG TGG CCA AAT AAT 1573
Val Cys Cys Val Asp Pro Ser Pro Leu Ser Met Trp Pro Asn Asn
    110                 115                 120
TAT GGT GTT TGG GTT GAT GAG TTT GAG AAT TTA GGA CTG GAA AAT 1618
Tyr Gly Val Trp Val Asp Glu Phe Glu Asn Leu Gly Leu Glu Asn
125                 130                 135
TGT TTA GAT CAT AAA TGG CCT ATG ACT TGT GTG CAT ATA AAT GAT 1663
Cys Leu Asp His Lys Trp Pro Met Thr Cys Val His Ile Asn Asp
            140                 145                 150
AAC AAA ACT AAG TAT TTG GGA AGA CCA TAT GGT AGA GTT AGT AGA 1708
Asn Lys Thr Lys Tyr Leu Gly Arg Pro Tyr Gly Arg Val Ser Arg
        155                 160                 165
AAG AAG CTG AAG TTG AAA TTG TTG AAT AGT TGT GTT GAG AAC AGA 1753
Lys Lys Leu Lys Leu Lys Leu Leu Asn Ser Cys Val Glu Asn Arg
    170                 175                 180
```

```
GTG AAG TTT TAT AAA GCT AAG GTT TGG AAA GTG GAA CAT GAA GAA 1798
Val Lys Phe Tyr Lys Ala Lys Val Trp Lys Val Glu His Glu Glu
                185                 190                 195
TTT GAG TCT TCA ATT GTT TGT GAT GAT GGT AAG AAG ATA AGA GGT 1843
Phe Glu Ser Ser Ile Val Cys Asp Asp Gly Lys Lys Ile Arg Gly
                200                 205                 210
AGT TTG GTT GTG GAT GCA AGT GGT TTT GCT AGT GAT TTT ATA GAG 1888
Ser Leu Val Val Asp Ala Ser Gly Phe Ala Ser Asp Phe Ile Glu
                215                 220                 225
TAT GAC AGG CCA AGA AAC CAT GGT TAT CAA ATT GCT CAT GGG GTT 1933
Tyr Asp Arg Pro Arg Asn His Gly Tyr Gln Ile Ala His Gly Val
                230                 235                 240
TTA GTA GAA GTT GAT AAT CAT CCA TTT GAT TTG GAT AAA ATG GTG 1978
Leu Val Glu Val Asp Asn His Pro Phe Asp Leu Asp Lys Met Val
                245                 250                 255
CTT ATG GAT TGG AGG GAT TCT CAT TTG GGT AAT GAG CCA TAT TTA 2023
Leu Met Asp Trp Arg Asp Ser His Leu Gly Asn Glu Pro Tyr Leu
                260                 265                 270
AGG GTG AAT AAT GCT AAA GAA CCA ACA TTC TTG TAT GCA ATG CCA 2068
Arg Val Asn Asn Ala Lys Glu Pro Thr Phe Leu Tyr Ala Met Pro
                275                 280                 285
TTT GAT AGA GAT TTG GTT TTC TTG GAA GAG ACT TCT TTG GTG AGT 2113
Phe Asp Arg Asp Leu Val Phe Leu Glu Glu Thr Ser Leu Val Ser
                290                 295                 300
CGT CCT GTT TTA TCG TAT ATG GAA GTA AAA AGA AGG ATG GTG GCA 2158
Arg Pro Val Leu Ser Tyr Met Glu Val Lys Arg Arg Met Val Ala
                305                 310                 315
AGA TTA AGG CAT TTG GGG ATC AAA GTG AAA AGT GTT ATT GAG GAA 2203
Arg Leu Arg His Leu Gly Ile Lys Val Lys Ser Val Ile Glu Glu
                320                 325                 330
GAG AAA TGT GTG ATC CCT ATG GGA GGA CCA CTT CCG CGG ATT CCT 2248
Glu Lys Cys Val Ile Pro Met Gly Gly Pro Leu Pro Arg Ile Pro
                335                 340                 345
CAA AAT GTT ATG GCT ATT GGT GGG AAT TCA GGG ATA GTT CAT CCA 2293
Gln Asn Val Met Ala Ile Gly Gly Asn Ser Gly Ile Val His Pro
                350                 355                 360
TCA ACA GGG TAC ATG GTG GCT AGG AGC ATG GCT TTA GCA CCA GTA 2338
Ser Thr Gly Tyr Met Val Ala Arg Ser Met Ala Leu Ala Pro Val
                365                 370                 375
CTA GCT GAA GCC ATC GTC GAG GGG CTT GGC TCA ACA AGA ATG ATA 2383
Leu Ala Glu Ala Ile Val Glu Gly Leu Gly Ser Thr Arg Met Ile
                380                 385                 390
AGA GGG TCT CAA CTT TAC CAT AGA GTT TGG AAT GGT TTG TGG CCT 2428
Arg Gly Ser Gln Leu Tyr His Arg Val Trp Asn Gly Leu Trp Pro
                395                 400                 405
TTG GAT AGA AGA TGT GTT AGA GAA TGT TAT TCA TTT GGG ATG GAG 2473
Leu Asp Arg Arg Cys Val Arg Glu Cys Tyr Ser Phe Gly Met Glu
                410                 415                 420
ACA TTG TTG AAG CTT GAT TTG AAA GGG ACT AGG AGA TTG TTT GAC 2518
Thr Leu Leu Lys Leu Asp Leu Lys Gly Thr Arg Arg Leu Phe Asp
                425                 430                 435
GCT TTC TTT GAT CTT GAT CCT AAA TAC TGG CAA GGG TTC CTT TCT 2563
Ala Phe Phe Asp Leu Asp Pro Lys Tyr Trp Gln Gly Phe Leu Ser
                440                 445                 450
TCA AGA TTG TCT GTC AAA GAA CTT GGT TTA CTC AGC TTG TGT CTT 2608
Ser Arg Leu Ser Val Lys Glu Leu Gly Leu Leu Ser Leu Cys Leu
                455                 460                 465
TTC GGA CAT GGC TCA AAC ATG ACT AGG TTG GAT ATT GTT ACA AAA 2653
Phe Gly His Gly Ser Asn Met Thr Arg Leu Asp Ile Val Thr Lys
                470                 475                 480
TGT CCT CTT CCT TTG GTT AGA CTG ATT GGC AAT CTA GCA ATA GAG 2698
Cys Pro Leu Pro Leu Val Arg Leu Ile Gly Asn Leu Ala Ile Glu
                485                 490                 495
AGC CTT TGA ATG TGA AAA GTT TGA ATC ATT TTC TTC ATT TTA ATT 2743
Ser Leu
    498
TCT TTG ATT ATT TTC ATA TTT TCT CAA TTG CAA AAG TGA GAT AAG 2788
AGC TAC ATA CTG TCA ACA AAT AAA CTA CTA TTG GAA AGT TAA AAT 2833
ATG TGT TTG TTG TAT GTT ATT CTA ATG GAA TGG ATT TTG TAA A   2876
```

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3265
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

```
ATC TCA TTG TAT AGC TTG TCT TTT GTT TCA GTC GTC TTA GGC TTG    45
GGT TAG TTG GTG TTG CTG TTT CAT ACT TCT ATC AAC CTT GTG TGA    90
GTT CCT TTA TAA AAT ATG ACT GTT GGA GGA AGT AAT TTA CCT TTA   135
GTT CGA CTA CAT CAA GAT TTG CAT CAT TCT CGT CCA AGA AAT CTT   180
AGT TTG AAG CCT TTT GGT CTG GTA TAT TTG TCA ATC TGA GCT TCG   225
CAA CTT TCT CAT GAC AGG GGT TTG TTG ACA TGC CTG ATT GTG CTC   270
TTC CTT TAC TTG ATA ATT GCT GCT TGT TGC GGA GGC ATC ACT CTA   315
CCT TCC TGC AGA TCA TGA ATT CTC TGA AAA GGA GCA CCA TAT TTG   360
CCG CAC TGT GGT TCA TAT TTC CAA TTA CAT TTA GAT GAA CTA TAT   405
CAT CAG GAG TGA AAG GTT ATT GTA TAA TCA ATC CAG TGG ATT CTC   450
GTT CTG GCA CCT TTA GAA GTA CAT GTG CGG AAA AGA ATG ATA AGG   495
TTT GTA TTG TTG TTG ACA AGG CCT GTT GCC TTT CTC ATT TGT AAA   540
TGT TCT GAA CGA CTC CTA AAT TAC TCT TAA AGT GTA AGG TCT TCC   585
GTG CCT GTT TGT ATA TAT AAT GCT GTG CCG TGA CTT ACC TTT TGT   630
ACC ATT TGT TCA AAT GTA TGG CCT GGA CAC TAG GGT TGT CAA AAA   675
TGT CTC ATG ACT TCA CCC TTC TTT CTT GTC TTG GTG CCC GTT TTA   720
TTG GTC TGA GAA CGG CGT GAT GCC AAA TTC TGC CGC TCC ACA GTG   765
AGC ATT TCG ATC TAC TGG AAA TTG ACC AAC TTA TTT TAT CAC TTG   810
ATA ACT AGA GTC TGG GTT CAA ACA AAA TCC AAT AAC TTC AAT CAT   855
ACA TTG TAT TTA TAT TGA AAA AAT TAT GCA CAA CTC AGT AAA TTA   900
CCT TTT TTT GCA GTC AAA AAT TCT AGA TCA GTT TCT AAT TAA TCA   945
AAA TGG CCT TTA TAG GGT CCC AGT TCC ATT AAT ATA CCT GCC GTC   990
CAT GCA CTG ATT ACA AGA CAA ATA CCT CAC TAT GTT TGT TAG TGC  1035
TTG GTA ATA TAA AAC CTT TTC TTT TAT GAG AAA GTT CAC CGA AAA  1080
TAA TTT TCT ATT TGT GGC ATA ACT AGT ATC GAA GTA TAT AGT GCA  1125
GAT TGA CAA GAA TTT AAT TTT GCA GTT GGG CAC ATG AAC AAT TTT  1170
CCT CAA AGT TGT AGA AAA TAT TTT TCA TTT TCT TGT CAC CGA AAA  1215
TTA TTT ATA ATT GAA ATT GAA ACC GAA TGA GCT GCA AGA CTC GAG  1260
TCG AAT TTC AAA AAA ATT GAC CAA CTA AAT ATG AAA AAA TCC GAA  1305
TAT ATC CCC CAC CCC CTA CCA AAC ACA TCC TAA AGT GAG GTA TAG  1350
ACT GGG ACT GGG ATT GGG AAA AGG GTA AAA TGC TTT CAC TAG CTT  1395
AGC AAA GAT TCC ACT TTG TTA GCT ATC TTT CTT TCT CAT TTC CTT  1440
TTT TCT TTT TCT TTT TTT TGT TAT ATA AGC CAA AGT AGG TAC CCA  1485
AAA GCA TCA ATA TTT TGT ATT GCT TGG TGA TTC CTC TTT ACT CCA  1530
GTA TTT CAT TTT CTA CAA GTT CCA CCT CCC TCC ATA ATT AAC CAT  1575
TAT CAA TCT TAT ACA TTT TCT ATA ATG GAA ACT CTT CTC AAG CCT  1620
                              Met Glu Thr Leu Leu Lys Pro
                                                5

TTT CCA TCT CTT TTA CTT TCC TCT CCT ACA CCC TAT AGG TCT ATT  1665
Phe Pro Ser Leu Leu Leu Ser Ser Pro Thr Pro Tyr Arg Ser Ile
        10                  15                  20

GTC CAA CAA AAT CCT TCT TTT CTA AGT CCC ACC ACC CAA AAA AAA  1710
Val Gln Gln Asn Pro Ser Phe Leu Ser Pro Thr Thr Gln Lys Lys
        25                  30                  35

TCA AGA AAA TGT CTT CTT AGA AAC AAA AGT AGT AAA CTT TTT TGT  1755
Ser Arg Lys Cys Leu Leu Arg Asn Lys Ser Ser Lys Leu Phe Cys
        40                  45                  50

AGC TTT CTT GAT TTA GCA CCC ACA TCA AAG CCA GAG TCT TTA AAT  1800
Ser Phe Leu Asp Leu Ala Pro Thr Ser Lys Pro Glu Ser Leu Asn
        55                  60                  65

GTT AAC ATC TCA TGG GTT GAT CCT AAT TCT GGT CGG GCT CAA TTC  1845
Val Asn Ile Ser Trp Val Asp Pro Asn Ser Gly Arg Ala Gln Phe
        70                  75                  80

GAC GTG ATC ATT ATC GGA GCT GGC CCT GCT GGG CTC AGG TTA GCT  1890
Asp Val Ile Ile Ile Gly Ala Gly Pro Ala Gly Leu Arg Leu Ala
        85                  90                  95

GAA CAA GTT TCT AAA TAT GGT ATT AAG GTA TGT TGT GTT GAC CCT  1935
Glu Gln Val Ser Lys Tyr Gly Ile Lys Val Cys Cys Val Asp Pro
       100                 105                 110

TCA CCA CTC TCC ATG TGG CCA AAT AAT TAT GGT GTT TGG GTT GAT  1980
Ser Pro Leu Ser Met Trp Pro Asn Asn Tyr Gly Val Trp Val Asp
       115                 120                 125

GAG TTT GAG AAT TTA GGA CTG GAA GAT TGT TTA GAT CAT AAA TGG  2025
Glu Phe Glu Asn Leu Gly Leu Glu Asp Cys Leu Asp His Lys Trp
       130                 135                 140

CCT ATG ACT TGT GTG CAT ATA AAT GAT AAC AAG ACT AAG TAT TTG  2070
```

```
Pro Met Thr Cys Val His Ile Asn Asp Asn Lys Thr Lys Tyr Leu
       145                 150                 155
GGA AGA CCA TAT GGT AGA GTT AGT AGA AAG AAG CTG AAG TTG AAA 2115
Gly Arg Pro Tyr Gly Arg Val Ser Arg Lys Lys Leu Lys Leu Lys
       160                 165                 170
TTG TTG AAC AGT TGT GTT GAG AAC AGA GTG AAG TTT TAT AAA GCT 2160
Leu Leu Asn Ser Cys Val Glu Asn Arg Val Lys Phe Tyr Lys Ala
       175                 180                 185
AAG GTT TGG AAA GTG GAA CAT GAA GAA TTT GAG TCT TCA ATT GTT 2205
Lys Val Trp Lys Val Glu His Glu Glu Phe Glu Ser Ser Ile Val
       190                 195                 200
TGT GAT GAT GGT AAG AAG ATA AGA GGT AGT TTG GTT GTG GAT GCA 2250
Cys Asp Asp Gly Lys Lys Ile Arg Gly Ser Leu Val Val Asp Ala
       205                 210                 215
AGT GGT TTT GCT AGT GAT TTT ATA GAG TAT GAC AAG CCA AGA AAC 2295
Ser Gly Phe Ala Ser Asp Phe Ile Glu Tyr Asp Lys Pro Arg Asn
       220                 225                 230
CAT GGT TAT CAA ATT GCT CAT GGG GTT TTA GTA GAA GTT GAT AAT 2340
His Gly Tyr Gln Ile Ala His Gly Val Leu Val Glu Val Asp Asn
       235                 240                 245
CAT CCA TTT GAT TTG GAT AAA ATG GTG CTT ATG GAT TGG AGG GAT 2385
His Pro Phe Asp Leu Asp Lys Met Val Leu Met Asp Trp Arg Asp
       250                 255                 260
TCT CAT TTA GGT AAT GAG CCA TAT TTA AGG GTG AAT AAT GCT AAA 2430
Ser His Leu Gly Asn Glu Pro Tyr Leu Arg Val Asn Asn Ala Lys
       265                 270                 275
GAA CCA ACA TTC TTG TAT GCA ATG CCA TTT GAT AGA AAT TTG GTT 2475
Glu Pro Thr Phe Leu Tyr Ala Met Pro Phe Asp Arg Asn Leu Val
       280                 285                 290
TTC TTG GAA GAG ACT TCT TTG GTG AGT CGT CCT GTG TTA TCG TAT 2520
Phe Leu Glu Glu Thr Ser Leu Val Ser Arg Pro Val Leu Ser Tyr
       295                 300                 305
ATG GAA GTA AAA AGA AGG ATG GTG GCA AGA TTA AGG CAT TTG GGG 2565
Met Glu Val Lys Arg Arg Met Val Ala Arg Leu Arg His Leu Gly
       310                 315                 320
ATC AAA GTG AGA AGT GTT ATT GAG GAA GAG AAA TGT GTG ATC CCT 2610
Ile Lys Val Arg Ser Val Ile Glu Glu Glu Lys Cys Val Ile Pro
       325                 330                 335
ATG GGA GGA CCA CTT CCG CGG ATT CCT CAA AAT GTT ATG GCT ATT 2655
Met Gly Gly Pro Leu Pro Arg Ile Pro Gln Asn Val Met Ala Ile
       340                 345                 350
GGT GGG AAT TCA GGG ATA GTT CAT CCA TCA ACG GGG TAC ATG GTG 2700
Gly Gly Asn Ser Gly Ile Val His Pro Ser Thr Gly Tyr Met Val
       355                 360                 365
GCT AGG AGC ATG GCT TTA GCA CCA GTA CTA GCT GAA GCC ATC GTC 2745
Ala Arg Ser Met Ala Leu Ala Pro Val Leu Ala Glu Ala Ile Val
       370                 375                 380
GAG GGG CTT GGC TCA ACA AGA ATG ATA AGA GGG TCT CAA CTT TAC 2790
Glu Gly Leu Gly Ser Thr Arg Met Ile Arg Gly Ser Gln Leu Tyr
       385                 390                 395
CAT AGA GTT TGG AAT GGT TTG TGG CCT TTG GAT AGA AGA TGT GTT 2835
His Arg Val Trp Asn Gly Leu Trp Pro Leu Asp Arg Arg Cys Val
       400                 405                 410
AGA GAA TGT TAT TCA TTT GGG ATG GAG ACA TTG TTG AAG CTT GAT 2880
Arg Glu Cys Tyr Ser Phe Gly Met Glu Thr Leu Leu Lys Leu Asp
       415                 420                 425
TTG AAA GGG ACT AGG AGA TTG TTT GAC GCT TTC TTT GAT CTT GAT 2925
Leu Lys Gly Thr Arg Arg Leu Phe Asp Ala Phe Phe Asp Leu Asp
       430                 435                 440
CCT AAA TAC TGG CAA GGG TTC CTT TCT TCA AGA TTG TCT GTC AAA 2970
Pro Lys Tyr Trp Gln Gly Phe Leu Ser Ser Arg Leu Ser Val Lys
       445                 450                 455
GAA CTT GGT TTA CTC AGC TTG TGT CTT TTC GGA CAT GGC TCA AAT 3015
Glu Leu Gly Leu Leu Ser Leu Cys Leu Phe Gly His Gly Ser Asn
       460                 465                 470
TTG ACT AGG TTG GAT ATT GTT ACA AAA TGT CCT GTT CCT TTG GTT 3060
Leu Thr Arg Leu Asp Ile Val Thr Lys Cys Pro Val Pro Leu Val
       475                 480                 485
AGA CTG ATT GGC AAT CTA GCA GTA GAG AGC CTT TGA ATG TGA AAA 3105
Arg Leu Ile Gly Asn Leu Ala Val Glu Ser Leu
       490                 495         498
GTT TGA ATC ATT TTC TTT ATT TTA ATT TCT TTG ATT ATT TTC ATA 3150
TTT TCT CAA TGC AAA AGT GAG AGA AGA CTA TAC ACT GTC AAC AAA 3195
TAA ACT ACT ATT GGA AAG TTA AAA TAA TGT GTG TGT TGT ATG TTA 3240
TGC TAA TGG AAT GGA TTG GTG TAA A                           3265
```

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1740

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

```
ATG GAA GCT CTT CTC AAG CCT TTT CCA TCT CTT TTA CTT TCC TCT   45
Met Glu Ala Leu Leu Lys Pro Phe Pro Ser Leu Leu Leu Ser Ser
  5                  10                  15
CCT ACA CCC TAT AGG TCT ATT GTC CAA CAA AAT CCT TCT TTT CTA   90
Pro Thr Pro Tyr Arg Ser Ile Val Gln Gln Asn Pro Ser Phe Leu
            20                  25                  30
AGT CCC ACC ACC AAA AAA AAT CAA GAA AAT GTC TTC TTA GAA ACA  135
Ser Pro Thr Thr Lys Lys Asn Gln Glu Asn Val Phe Leu Glu Thr
        35                  40                  45
AAA GTA GTA AAC TTT TTT GTA GCT TTC TTG ATT TAG CAC CCA CAT  180
Lys Val Val Asn Phe Phe Val Ala Phe Leu Ile
    50                  55  56
CAA AGC CAG AGT CTT TAA ATG TTA ACA TCT CAT GGG TTG ATC CTA  225
ATT CGA ATC GGG CTC AAT TCG ACG TGA TCA TTA TCG GAG CTG GCC  270
CTG CTG GGC TCA GGC TAG CTG AAC AAG TTT CTA AAT ATG GTA TTA  315
AGG TAT GTT GTG TTG ACC CTT CAC CAC TCT CCA TGT GGC CAA ATA  360
ATT ATG GTG TTT GGG TTG ATG AGT TTG AGA ATT TAG GAC TGG AAA  405
ATT GTT TAG ATC ATA AAT GGC CTA TGA CTT GTG TGC ATA TAA ATG  450
ATA ACA AAA CTA AGT ATT TGG GAA GAC CAT ATG GTA GAG TTA GTA  495
GAA AGA AGC TGA AGT TGA AAT TGT TGA ATA GTT GTG TTG AGA ACA  540
GAG TGA AGT TTT ATA AAG CTA AGG TTT GGA AAG TGG AAC ATG AAG  585
AAT TTG AGT CTT CAA TTG TTT GTG ATG ATG GTA AGA AGA TAA GAG  630
GTA GTT TGG TTG TGG ATG CAA GTG GTT TTG CTA GTG ATT TTA TAG  675
AGT ATG ACA GGC CAA GAA ACC ATG GTT ATC AAA TTG CTC ATG GGG  720
TTT TAG TAG AAG TTG ATA ATC ATC CAT TTG ATT TGG ATA AAA TGG  765
TGC TTA TGG ATT GGA GGG ATT CTC ATT TGG GTA ATG AGC CAT ATT  810
TAA GGG TGA ATA ATG CTA AAG AAC CAA CAT TCT TGT ATG CAA TGC  855
CAT TTG ATA GAG ATT TGG TTT TCT TGG AAG AGA CTT CTT TGG TGA  900
GTC GTC CTG TGT TAT CGT ATA TGG AAG TAA AAA GAA GGA TGG TGG  945
CAA GAT TAA GGC ATT TGG GGA TCA AAG TGA AAA GTG TTA TTG AGG  990
AAG AGA AAT GTG TGA TCC CTA TGG GAG GAC CAC TTC CGC GGA TTC 1035
CTC AAA ATG TTA TGG CTA TTG GTG GGA ATT CAG GGA TAG TTC ATC 1080
CAT CAA CAG GGT ACA TGG TGG CTA GGA GCA TGG CTT TAG CAC CAG 1125
TAC TAG CTG AAG CCA TCG TCG AGG GGC TTG GCT CAA CAA GAA TGA 1170
TAA GAG GGT CTC AAC TTT ACC ATA GAG TTT GGA ATG GTT TGT GGC 1215
CTT TGG ATA GAA GAT GTG TTA GAG AAT GTT ATT CAT TTG GGA TGG 1260
AGA CAT TGT TGA AGC TTG ATT TGA AAG GGA CTA GGA GAT TGT TTG 1305
ACG CTT TCT TTG ATC TTG ATC CTA AAT ACT GGC AAG GGT TCC TTT 1350
CTT CAA GAT TGT CTG TCA AAG AAA CTT GGT TTA CTC AGC TTG TGT 1395
CTT TTC GGA CAT GGC TCA AAC ATG ACT AGG TTG GGA TAT TGT TAC 1440
AAA ATG TCC TCT TCC TTT GGT TAG ACT GAT TGG CAA TCT AGC AAT 1485
AGA GAG CCT TTG AAA TGT GAA AAG TTT GAA TCA TTT TCT TCA TTT 1530
TAA TTT CTT TGA TTA TTT TCA TAT TTT CTC AAT TGC AGA ATG AGA 1575
TAA AAA CTA CAT ACT GTC GAC AAA TAA ACT ACT ATT GGA ANG TTA 1620
AAA TAA TGT GTG TGT TGN ATG TTA NGC CTA ATG GAA NGG ATG NGG 1665
TTA NGC AAT TTA TGA ACT GNN CGC TCT GTT CGC TTA AAA NCC TTG 1710
GTT CCA CCT TAA NGG AAN GGN CCG GCC ATT                     1740
```

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 498

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

```
Met Glu Ala Leu Leu Lys Pro Phe Pro Ser Leu Leu Leu Ser Ser
  5                  10                  15
Pro Thr Pro His Arg Ser Ile Phe Gln Gln Asn Pro Ser Phe Leu
            20                  25                  30
Ser Pro Thr Thr Lys Lys Lys Ser Arg Lys Cys Leu Leu Arg Asn
        35                  40                  45
Lys Ser Ser Lys Leu Phe Cys Ser Phe Leu Asp Leu Ala Pro Thr
```

```
                50                  55                  60
Ser Lys Pro Glu Ser Leu Asp Val Asn Ile Ser Trp Val Asp Pro
                65                  70                  76
Asn Ser Asn Arg Ala Gln Phe Asp Val Ile Ile Ile Gly Ala Gly
                80                  85                  90
Pro Ala Gly Leu Arg Leu Ala Glu Gln Val Ser Lys Tyr Gly Ile
                95                 100                 105
Lys Val Cys Cys Val Asp Pro Ser Pro Leu Ser Met Trp Pro Asn
               110                 115                 120
Asn Tyr Gly Val Trp Val Asp Glu Phe Glu Asn Leu Gly Leu Glu
               125                 130                 135
Asn Cys Leu Asp His Lys Trp Pro Met Thr Cys Val His Ile Asn
               140                 145                 150
Asp Asn Lys Thr Lys Tyr Leu Gly Arg Pro Tyr Gly Arg Val Ser
               155                 160                 165
Arg Lys Lys Leu Lys Leu Lys Leu Leu Asn Ser Cys Val Glu Asn
               170                 175                 180
Arg Val Lys Phe Tyr Lys Ala Lys Val Trp Lys Val Glu His Glu
               185                 190                 195
Glu Phe Glu Ser Ser Ile Val Cys Asp Asp Gly Lys Lys Ile Arg
               200                 205                 210
Gly Ser Leu Val Val Asp Ala Ser Gly Phe Ala Ser Asp Phe Ile
               215                 220                 225
Glu Tyr Asp Arg Pro Arg Asn His Gly Tyr Gln Ile Ala His Gly
               230                 235                 240
Val Leu Val Glu Val Asp Asn His Pro Phe Asp Leu Asp Lys Met
               245                 250                 255
Val Leu Met Asp Trp Arg Asp Ser His Leu Gly Asn Glu Pro Tyr
               260                 265                 270
Leu Arg Val Asn Asn Ala Lys Glu Pro Thr Phe Leu Tyr Ala Met
               275                 280                 285
Pro Phe Asp Arg Asp Leu Val Phe Leu Glu Glu Thr Ser Leu Val
               290                 295                 300
Ser Arg Pro Val Leu Ser Tyr Met Glu Val Lys Arg Arg Met Val
               305                 310                 315
Ala Arg Leu Arg His Leu Gly Ile Lys Val Lys Ser Val Ile Glu
               320                 325                 330
Glu Glu Lys Cys Val Ile Pro Met Gly Gly Pro Leu Pro Arg Ile
               335                 340                 345
Pro Gln Asn Val Met Ala Ile Gly Gly Asn Ser Gly Ile Val His
               350                 355                 360
Pro Ser Thr Gly Tyr Met Val Ala Arg Ser Met Ala Leu Ala Pro
               365                 370                 375
Val Leu Ala Glu Ala Ile Val Glu Gly Leu Gly Ser Thr Arg Met
               380                 385                 390
Ile Arg Gly Ser Gln Leu Tyr His Arg Val Trp Asn Gly Leu Trp
               395                 400                 405
Pro Leu Asp Arg Arg Cys Val Arg Glu Cys Tyr Ser Phe Gly Met
               410                 415                 420
Glu Thr Leu Leu Lys Leu Asp Leu Lys Gly Thr Arg Arg Leu Phe
               425                 430                 435
Asp Ala Phe Phe Asp Leu Asp Pro Lys Tyr Trp Gln Gly Phe Leu
               440                 445                 450
Ser Ser Arg Leu Ser Val Lys Glu Leu Gly Leu Leu Ser Leu Cys
               455                 460                 465
Leu Phe Gly His Gly Ser Asn Met Thr Arg Leu Asp Ile Val Thr
               470                 475                 480
Lys Cys Pro Leu Pro Leu Val Arg Leu Ile Gly Asn Leu Ala Ile
               485                 490                 495
Glu Ser Leu
       498
```

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 498

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

```
Met Glu Ala Leu Leu Lys Pro Phe Pro Ser Leu Leu Leu Ser Ser
                 5                  10                  15
Pro Thr Pro His Arg Ser Ile Phe Gln Gln Asn Pro Ser Phe Leu
```

```
                20                  25                  30
Ser Pro Thr Thr Lys Lys Lys Ser Arg Lys Cys Leu Leu Arg Asn
                35                  40                  45
Lys Ser Ser Lys Leu Phe Cys Ser Phe Leu Asp Leu Ala Pro Thr
                50                  55                  60
Ser Lys Pro Glu Ser Leu Asp Val Asn Ile Ser Trp Val Asp Pro
                65                  70                  76
Asn Ser Asn Arg Ala Gln Phe Asp Val Ile Ile Ile Gly Ala Gly
                80                  85                  90
Pro Ala Gly Leu Arg Leu Ala Glu Gln Val Ser Lys Tyr Gly Ile
                95                  100                 105
Lys Val Cys Cys Val Asp Pro Ser Pro Leu Ser Met Trp Pro Asn
                110                 115                 120
Asn Tyr Gly Val Trp Val Asp Glu Phe Glu Asn Leu Gly Leu Glu
                125                 130                 135
Asn Cys Leu Asp His Lys Trp Pro Met Thr Cys Val His Ile Asn
                140                 145                 150
Asp Asn Lys Thr Lys Tyr Leu Gly Arg Pro Tyr Gly Arg Val Ser
                155                 160                 165
Arg Lys Lys Leu Lys Leu Lys Leu Leu Asn Ser Cys Val Glu Asn
                170                 175                 180
Arg Val Lys Phe Tyr Lys Ala Lys Val Trp Lys Val Glu His Glu
                185                 190                 195
Glu Phe Glu Ser Ser Ile Val Cys Asp Asp Gly Lys Lys Ile Arg
                200                 205                 210
Gly Ser Leu Val Val Asp Ala Ser Gly Phe Ala Ser Asp Phe Ile
                215                 220                 225
Glu Tyr Asp Arg Pro Arg Asn His Gly Tyr Gln Ile Ala His Gly
                230                 235                 240
Val Leu Val Glu Val Asp Asn His Pro Phe Asp Leu Asp Lys Met
                245                 250                 255
Val Leu Met Asp Trp Arg Asp Ser His Leu Gly Asn Glu Pro Tyr
                260                 265                 270
Leu Arg Val Asn Asn Ala Lys Glu Pro Thr Phe Leu Tyr Ala Met
                275                 280                 285
Pro Phe Asp Arg Asp Leu Val Phe Leu Glu Glu Thr Ser Leu Val
                290                 295                 300
Ser Arg Pro Val Leu Ser Tyr Met Glu Val Lys Arg Arg Met Val
                305                 310                 315
Ala Arg Leu Arg His Leu Gly Ile Lys Val Lys Ser Val Ile Glu
                320                 325                 330
Glu Glu Lys Cys Val Ile Pro Met Gly Gly Pro Leu Pro Arg Ile
                335                 340                 345
Pro Gln Asn Val Met Ala Ile Gly Gly Asn Ser Gly Ile Val His
                350                 355                 360
Pro Ser Thr Gly Tyr Met Val Ala Arg Ser Met Ala Leu Ala Pro
                365                 370                 375
Val Leu Ala Glu Ala Ile Val Glu Gly Leu Gly Ser Thr Arg Met
                380                 385                 390
Ile Arg Gly Ser Gln Leu Tyr His Arg Val Trp Asn Gly Leu Trp
                395                 400                 405
Pro Leu Asp Arg Arg Cys Val Arg Glu Cys Tyr Ser Phe Gly Met
                410                 415                 420
Glu Thr Leu Leu Lys Leu Asp Leu Lys Gly Thr Arg Arg Leu Phe
                425                 430                 435
Asp Ala Phe Phe Asp Leu Asp Pro Lys Tyr Trp Gln Gly Phe Leu
                440                 445                 450
Ser Ser Arg Leu Ser Val Lys Glu Leu Gly Leu Leu Ser Leu Cys
                455                 460                 465
Leu Phe Gly His Gly Ser Asn Met Thr Arg Leu Asp Ile Val Thr
                470                 475                 480
Lys Cys Pro Leu Pro Leu Val Arg Leu Ile Gly Asn Leu Ala Ile
                485                 490                 495
Glu Ser Leu
        498
```

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 498

(B) TYPE: amino acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

```
Met Glu Thr Leu Leu Lys Pro Phe Pro Ser Leu Leu Leu Ser Ser
 1               5                  10                  15
Pro Thr Pro Tyr Arg Ser Ile Val Gln Gln Asn Pro Ser Phe Leu
            20                  25                  30
Ser Pro Thr Thr Gln Lys Lys Ser Arg Lys Cys Leu Leu Arg Asn
        35                  40                  45
Lys Ser Ser Lys Leu Phe Cys Ser Phe Leu Asp Leu Ala Pro Thr
    50                  55                  60
Ser Lys Pro Glu Ser Leu Asn Val Asn Ile Ser Trp Val Asp Pro
65                  70                  76
Asn Ser Gly Arg Ala Gln Phe Asp Val Ile Ile Ile Gly Ala Gly
            80                  85                  90
Pro Ala Gly Leu Arg Leu Ala Glu Gln Val Ser Lys Tyr Gly Ile
        95                  100                 105
Lys Val Cys Cys Val Asp Pro Ser Pro Leu Ser Met Trp Pro Asn
    110                 115                 120
Asn Tyr Gly Val Trp Val Asp Glu Phe Glu Asn Leu Gly Leu Glu
    125                 130                 135
Asp Cys Leu Asp His Lys Trp Pro Met Thr Cys Val His Ile Asn
    140                 145                 150
Asp Asn Lys Thr Lys Tyr Leu Gly Arg Pro Tyr Gly Arg Val Ser
    155                 160                 165
Arg Lys Lys Leu Lys Leu Lys Leu Leu Asn Ser Cys Val Glu Asn
    170                 175                 180
Arg Val Lys Phe Tyr Lys Ala Lys Val Trp Lys Val Glu His Glu
    185                 190                 195
Glu Phe Glu Ser Ser Ile Val Cys Asp Asp Gly Lys Lys Ile Arg
    200                 205                 210
Gly Ser Leu Val Val Asp Ala Ser Gly Phe Ala Ser Asp Phe Ile
    215                 220                 225
Glu Tyr Asp Lys Pro Arg Asn His Gly Tyr Gln Ile Ala His Gly
    230                 235                 240
Val Leu Val Glu Val Asp Asn His Pro Phe Asp Leu Asp Lys Met
    245                 250                 255
Val Leu Met Asp Trp Arg Asp Ser His Leu Gly Asn Glu Pro Tyr
    260                 265                 270
Leu Arg Val Asn Asn Ala Lys Glu Pro Thr Phe Leu Tyr Ala Met
    275                 280                 285
Pro Phe Asp Arg Asn Leu Val Phe Leu Glu Glu Thr Ser Leu Val
    290                 295                 300
Ser Arg Pro Val Leu Ser Tyr Met Glu Val Lys Arg Arg Met Val
    305                 310                 315
Ala Arg Leu Arg His Leu Gly Ile Lys Val Arg Ser Val Ile Glu
    320                 325                 330
Glu Glu Lys Cys Val Ile Pro Met Gly Gly Pro Leu Pro Arg Ile
    335                 340                 345
Pro Gln Asn Val Met Ala Ile Gly Gly Asn Ser Gly Ile Val His
    350                 355                 360
Pro Ser Thr Gly Tyr Met Val Ala Arg Ser Met Ala Leu Ala Pro
    365                 370                 375
Val Leu Ala Glu Ala Ile Val Glu Gly Leu Gly Ser Thr Arg Met
    380                 385                 390
Ile Arg Gly Ser Gln Leu Tyr His Arg Val Trp Asn Gly Leu Trp
    395                 400                 405
Pro Leu Asp Arg Arg Cys Val Arg Glu Cys Tyr Ser Phe Gly Met
    410                 415                 420
Glu Thr Leu Leu Lys Leu Asp Leu Lys Gly Thr Arg Arg Leu Phe
    425                 430                 435
Asp Ala Phe Phe Asp Leu Asp Pro Lys Tyr Trp Gln Gly Phe Leu
    440                 445                 450
Ser Ser Arg Leu Ser Val Lys Glu Leu Gly Leu Leu Ser Leu Cys
    455                 460                 465
Leu Phe Gly His Gly Ser Asn Leu Thr Arg Leu Asp Ile Val Thr
    470                 475                 480
Lys Cys Pro Val Pro Leu Val Arg Leu Ile Gly Asn Leu Ala Val
    485                  90                 495
Glu Ser Leu
    498
```

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 56

(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Met Glu Ala Leu Leu Lys Pro Phe Pro Ser Leu Leu Leu Ser Ser
5 10 15
Pro Thr Pro Tyr Arg Ser Ile Val Gln Gln Asn Pro Ser Phe Leu
20 25 30
Ser Pro Thr Thr Lys Lys Asn Gln Glu Asn Val Phe Leu Glu Thr
35 40 45
Lys Val Val Asn Phe Phe Val Ala Phe Leu Ile
50 55

(2) INFORMATION FOR SEQ ID NO:21:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 nucleic acids
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

TGACTTCACC CTTCTTTCTT GTCTTC 26

(2) INFORMATION FOR SEQ ID NO:22:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 13 nucleic acids
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

AGAGTCTGGG TTC 13

(2) INFORMATION FOR SEQ ID NO:23:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 9 nucleic acids
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CTAGTATCG 9

(2) INFORMATION FOR SEQ ID NO:24:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 nucleic acids
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CTAAATAT 8

(2) INFORMATION FOR SEQ ID NO:25:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 nucleic acids
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

AATTTTCAAA 10

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/18327

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :Please See Extra Sheet.
US CL :Please See Extra Sheet.
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/6, 243, 252.3, 419, 468, 471; 536/23.2, 23.6, 24.5; 800/298

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, STN, AGRICOLA, CAPLUS, BIOSIS, EMBASE
search terms: lycopene cyclase, DNA, cDNA, gene, nucleic acid, antisense, plant, tomato, Lycopersicon, carotene

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category°	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X - Y	WO 96/36717 A1 (CENTRE NATIONAL DE LA RECHERCHE SCIENTIFIQUE) 21 November 1996, see entire patent.	1,2,6-12,14 -17,23 ——— 3,13
Y	COOK, P.D. Medicinal Chemistry of Antisense Oligonucleotides - Future Opportunities. Anti-Cancer Drug Design. 1991, Vol. 6, pages 585-607, see entire article.	13

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

°	Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A"	document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E"	earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L"	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O"	document referring to an oral disclosure, use, exhibition or other means		
"P"	document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search
19 OCTOBER 1999

Date of mailing of the international search report
05 NOV 1999

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231
Facsimile No. (703) 305-3230

Authorized officer
AMY NELSON
Telephone No. (703) 308-0196

Form PCT/ISA/210 (second sheet)(July 1992)☆

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/18327

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☒ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
1-4, 6-22, 24-25

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest.

☒ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/18327

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (6):

A01H 5/00; C12N 1/00, 1/21, 5/14, 15/29, 15/52, 15/70, 15/74, 15/82; C12Q 1/68

A. CLASSIFICATION OF SUBJECT MATTER:
US CL :

435/6, 243, 252.3, 419, 468, 471; 536/23.2, 23.6, 24.5; 800/298

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING
This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s) 1-4,6-9, drawn to DNA and transformed host cell.
Group II, claim(s) 5, drawn to protein.
Group III, claim(s) 10-17, drawn to antisense method.
Group IV, claim(s) 18-22,24,25, drawn to promoter construct, transformed host cell, and transgenic plant.
Group V, claim(s) 23, drawn to hybridization method.

The inventions listed as Groups I, II, III, IV, and V do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons:

Claim 1 is indefinite due to the claim language "variants," and the claims of Group I read on previously disclosed isolated DNAs encoding lycopene cyclase, such as those disclosed by Kuntz *et al.* (WO96/36717). Therefore, there is no special technical feature under PCT Rule 13.2, which links the DNA of Group I, the protein of Group II, the antisense method of Group III and the hybridization method of Group V.

Furthermore, the promoter construct, and cells and plants transformed therewith, of Group IV are structurally and functionally distinct from the coding DNA of Group I, and therefore the coding DNA of Group I and the promoter construct of Group IV are not linked by a special technical feature under PCT Rule 13.2.

Hence, the inventions of Groups I, II, III, IV, and V do not relate to a single inventive concept under PCT Rule 13.1.